

Role of dietary lipids and gut microbiome-derived lipids in regulation of intestinal homeostasis and modulation of inflammatory diseases

Chi Yan[†], Shou-He Huang[†], Huafang Ding, Wen-Sen He, Hanyue Zhu and Zhen-Yu Chen^{*}

School of Life Sciences, Faculty of Science, The Chinese University of Hong Kong, Shatin, NT, Hong Kong, China

[†]These authors contributed equally to this work.

^{*}**Corresponding author:** Zhen-Yu Chen, School of Life Sciences, Faculty of Science, The Chinese University of Hong Kong, Shatin, NT, Hong Kong, China. E-mail: zhenyuchen@cuhk.edu.hk

DOI: 10.26599/JFB.2024.95028391

Received: December 13, 2024; **Revised received & accepted:** December 20, 2024

Citation: Yan, C., Huang, S.-H., Ding, H., He, W.-S., Zhu, H., and Chen, Z.-Y. (2024). Role of dietary lipids and gut microbiome-derived lipids in regulation of intestinal homeostasis and modulation of inflammatory diseases. *J. Food Bioact.* 28: 1–23.

Abstract

Gut health is the foundation of overall health. Inflammatory bowel disease (IBD) including Crohn's disease and ulcerative colitis is a kind of chronic relapsing and idiopathic immune dysbiosis in the intestinal tract. Recent studies have highlighted the effects of various dietary components on the progress of IBD. Mounting evidence has demonstrated that both dietary lipids and gut microbiome-derived lipids play a crucial role in gut health. They can directly or indirectly change the composition of gut microbiota, modulate the metabolism of colonic epithelial cells, influence the integrity of gut barrier and regulate the immune function. This review aims to define the key classes of dietary lipids and microbial-derived lipids, elucidate the interaction of these lipids with gut microbiota, discuss their effects on the intestinal homeostasis, and provide the future perspective in the research of gut health.

Keywords: Inflammatory bowel disease (IBD); Gut microbiota; Dietary lipids; Gut microbiome-derived lipids; Gut dysbiosis.

1. Introduction

The gastrointestinal (GI) tract is the largest immune organ in humans, consisting of complicated networks of various immune cells and epithelial cells (Kaur *et al.*, 2023). Intestinal homeostasis is the foundation of overall health in humans. Intestine-related inflammation can trigger a variety of systemic diseases including inflammatory bowel disease (IBD) and colorectal cancer (Jia, *et al.*, 2020b). IBD, such as Crohn's disease and ulcerative colitis, is a kind of chronic relapsing and idiopathic immune dysbiosis in GI tracts, accompanied by dysregulation of immune responses, causing a large number of destructive inflammatory cell infiltration and excessive production of pro-inflammatory factors (Lloyd-Price *et al.*, 2019). Recent epidemiological studies have clearly demonstrated that the incidence of IBD is substantially increasing (Chen

et al., 2023a). The number of patients with IBD has risen from 3.7 to more than 6.8 million from 1990 to 2017 around the world (Alatab *et al.*, 2020), demonstrating a need in management of GI health. Although numerous investigations have been performed to study the etiology of IBD, its underlying mechanisms remain unclear. It is hypothesized that IBD is associated with susceptibility of gene loci, environmental risk factors, and gut microbiota composition (Kobayashi *et al.*, 2020; Wu *et al.*, 2021).

Numerous therapies and medicines are available to treat IBD. Mesalazine (decreasing the expression of pro-inflammatory factors), corticosteroid (suppressing the activation of immune systems), biological therapy (fecal microbiota transplantation), surgical management (removing the damaged intestinal tissue), and immunosuppression (regulating the amount of relevant immune cells) have been widely used to alleviate inflammatory symptoms

of IBD, but the treatments outcomes are still challenging with some serious and unexpected side-effects, for example, medicine dependence, liver and kidney toxicity (Chen et al., 2023b; Hartwig et al., 2021). In addition, there are also some other limitations for current IBD treatments. For example, the application of fecal microbiota transfer, which helps to re-establish gut microbial communities of patients, is still controversial due to its transient outcomes, potential pathogenic infections, and unstable reproducibility (Pigneur and Sokol, 2016). With the advances in medical development, some novel concepts and strategies have been proposed to enable physicians to have better approaches for IBD management.

Recent studies have also highlighted the crucial role of various dietary components in the progress of IBD (Ananthkrishnan, 2015). Lipids are one of the three largest nutrients for humans (Hosomi et al., 2020). Intake of lipids can cause both beneficial and harmful effects in humans, for example, dietary fat is required for humans as the major energy income, however, its excessive intake can cause numerous chronic diseases, such as atherosclerosis, obesity, and even cancer. After consumption, dietary lipids are first digested in the small intestine and approximately 95% of lipids will be absorbed in the GI tracts (Armand, 2007). Numerous studies have indicated that dietary lipids have significant effects on intestinal homeostasis and are related to various inflammatory diseases, including the regulation of the colonic physical barrier and immune systems (Okamura et al., 2021; Rohr et al., 2020). Furthermore, dietary lipids can exert various effects on gut health by modulating the composition of gut microbiota (Ye et al., 2021).

Gut microbiota-derived molecules have demonstrated to be one of the important factors that can affect the physiological conditions of the host (Koppel et al., 2017). The effect of lipids derived from gut microbiota on host health has been understudied (Eichelmann et al., 2022). Recent studies have shown that gut microbiota-derived lipids can affect the homeostasis in GI tracts and modulate the inflammatory diseases (Brown et al., 2023). The bacterial-derived lipids can directly stimulate the metabolism of colonic epithelial cells and improve the integrity of gut barrier (Kelly et al., 2015). In addition, gut microbiota can also produce various kinds of lipids that have been proven as potential factors to regulate immune systems, such as short-chain fatty acids (SCFAs) and phospholipids (Cai et al., 2022; Yao et al., 2022).

There are some research gaps about the influence of dietary and gut microbiota-derived lipids on gut health. For example, some functional edible oils have been applied to improve the symptoms of IBD, but the discrete roles of different lipid components in IBD have not been well understood (Prabha et al., 2023). Many researchers have focused on the functionality of gut microbiota in health, but the important roles of lipids from the gut microbiota membrane are ignored. This review aims to highlight and update the biological effects of different dietary lipids on gut microbiota in the regulation of intestinal homeostasis and IBD. The potential underlying mechanisms of lipids in the modulating of gut health via gut microbiome-dependent or gut microbiome-independent manners will be discussed.

2. Gut homeostasis and mechanisms of IBD

The gut homeostasis is a complicated equilibrium maintained by a large number of factors. Beginning from the surface of the GI tract, the crucial factor is the gut bacteria which can educate immune systems and maintain gut barrier functions. A healthy intestinal condition can prevent the infiltration of bacteria across the

gut barrier and establish immune tolerance toward gut microbiota (Zheng et al., 2020). The intestinal homeostasis and health mainly depend on complicated interactions among the gut microbiome, intestinal barrier, and host immune systems (Maloy and Powrie, 2011). Although the pathophysiology of IBD is multifaceted and is not fully elucidated, current research allows us to summarize some main components that trigger and contribute to the incidence of intestinal inflammation: gut microbiota, barrier, and immune systems (Figure 1).

2.1. Gut microbiota

The GI tract is home to trillions of bacteria that perform diverse roles in human health. Studies have declared that numerous gut microbiota could be linked with the gene expression in hosts (Alexander et al., 2022; Venema, 2010). There is adequate evidence that the offspring can obtain gut microbiota from mothers in many ways (Yang et al., 2021). The environmental factors that come after birth, including dietary components, medicines, and even sleeping models can exert crucial effects on modulating the composition of gut microbiota in fetuses, whilst the composition of gut microbiota can be more stable in adults even though it can still be affected by various factors (Dominguez et al., 2011). Extensive studies based on antibiotics-treated mice or germ-free mice have revealed the indispensable roles of gut microbiota in human development and health. In normal conditions-raised mice colonized with the conventional microbial community, some dietary factors especially some complex carbohydrates can also be further digested and broken by bacteria and then provide nutrients that can nourish locally or systemically to the host (Oliphant and Allen-Vercoe, 2019). In contrast to wild-type mice, the cecum of germ-free mice is enlarged by four to eightfold due to the accumulation of undigested fibers and intestinal transit is also prolonged (Manca et al., 2020). In addition to dietary factors digestion as afore-described, the defective gut microbiota in germ-free mice can also induce a decrease of intestinal barrier integrity, a thinner mucus layer in the colon part, through decreasing Muc2 expression and secretion in the proximal colon (Bergstrom et al., 2020). Immune systems are also impacted in germ-free mice, for example, germ-free mice have hypoplastic Peyer's patches and diminished IgA-producing and CD4⁺ T cells in the intestine (Lin and Zhang, 2017).

Dysbiosis of gut microbiota has been related to the occurrence of IBD at the clinical level (Chassaing and Darfeuille-Michaud, 2011; de Souza and Fiocchi, 2016). In patients suffering from Crohn's disease, the relative abundance of *Bacteroidetes* and *Proteobacteria* increased, whereas *Firmicutes* was reduced compared with healthy individuals (Man et al., 2011). Meanwhile, some studies have observed that gut microbiota abnormalities in Crohn's disease are more significant than ulcerative colitis (Qiu et al., 2022). A decrease in *Roseburia hominis* and *Faecalibacterium prausnitzii* was reported in ulcerative colitis patients (Machiels et al., 2014). The pathogenic roles of some bacteria have also been evaluated in animal models, such as *Eggerthella lenta* which is enriched in IBD patients and exacerbated dextran sodium sulfate (DSS)-induced colitis in mice via inducing the activation of Th17 cells in antigen-independent mechanisms (Alexander et al., 2022). In addition, the decrease of some crucial metabolisms from gut microbiota was also reported, for example, SCFAs which have been connected with gut barrier functions and immune system regulation based on various murine models (Furusawa et al., 2013; Kelly et al., 2015). Remarkably, some studies showed that improvement of IBD could be achieved in some patients via fecal microbiota transplantation (Moayyedi et al., 2015; Paramsothy et al., 2017).

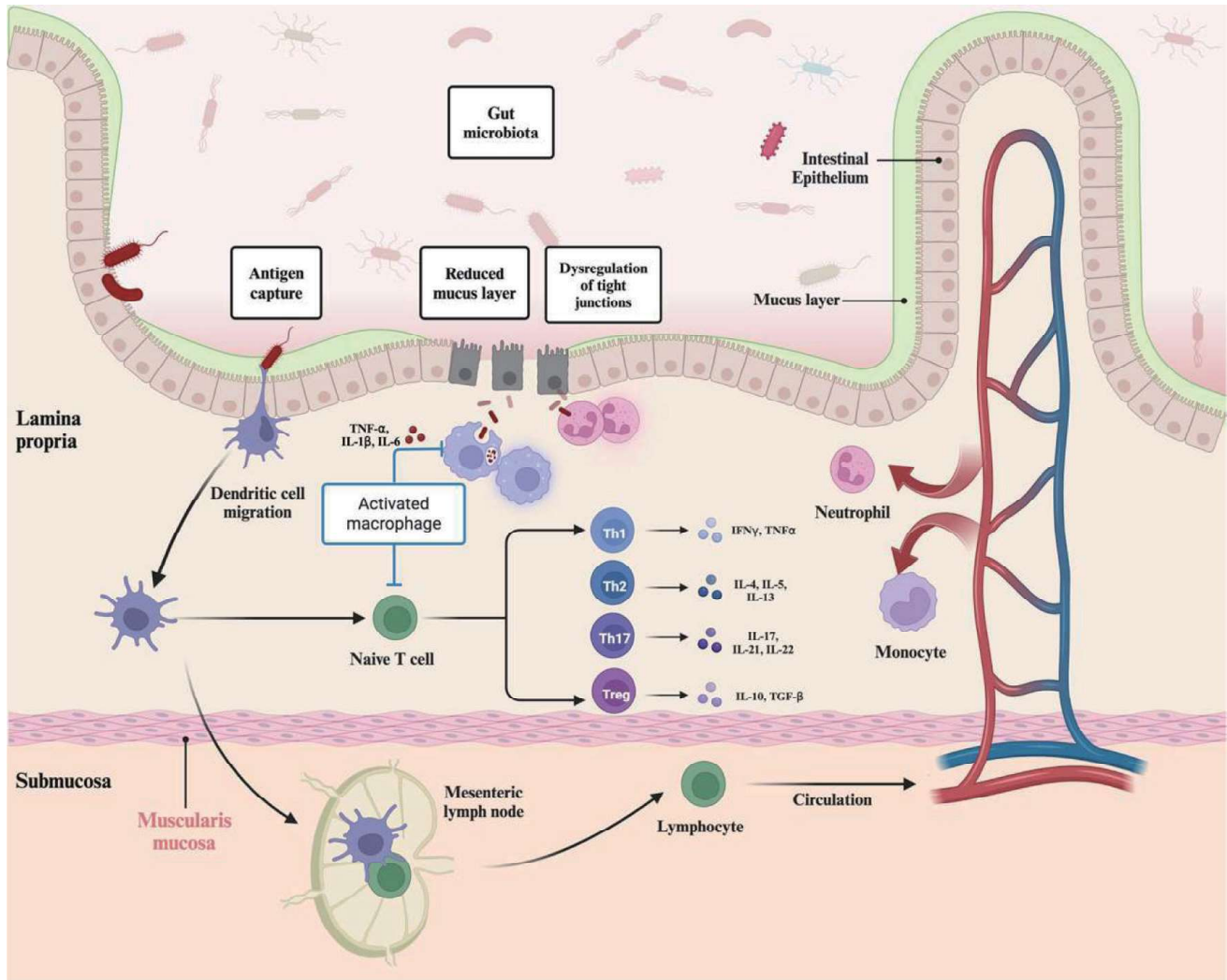


Figure 1. Dysbiosis and pathophysiology of inflammatory bowel disease (IBD). Multiple associated pathways, including gut microbiota, intestinal barrier, and immune systems, contribute to the incidence of IBD. In homeostasis conditions, trillions of commensal bacteria in the gastrointestinal tracts can help to mature the immune systems and produce beneficial metabolites, such as short-chain fatty acids. In IBD conditions, the composition of gut microbiota and metabolic profiles are altered, accompanied by an increase in some harmful bacteria, such as *Escherichia coli*. Impairment of the mucus layer and tight junction proteins permit the infiltration of microbiota and related antigens in lamina propria, activating innate immune systems and antigen-presenting cells to recruit neutrophils as the first responders in IBD. Neutrophils infiltrate and contribute to the first immune line via constructing neutrophil extracellular traps, excessive antigens can be recognized by macrophages and dendritic cells to further stimulate the activation of the adaptive immune system. With the stimulation and binding with antigen-presenting cells, naïve T cells can polarize and differentiate into various T helper and regulator cells, including Th1, Th2, Th17, and Treg, further contributing to the progress of IBD with the production of numerous cytokines. Figure 1 was created in Biorender.com (Agreement number: QL27CARACU).

Although this transplantation treatment is not sufficient for some patients to cure IBD and the therapy outcome is still challenging to predict, the important roles of gut microbiota in IBD have been revealed with this evidence.

With the increasing evidence and studies about the influence of gut microbiota on IBD, there are also emerging questions that still limit the application of related therapy in IBD management. Evidence exists that the function and roles of bacteria can be different in IBD progress. A prior study demonstrated that the relative abundance of *Akkermansia muciniphila* significantly decreased in TLR4^{-/-} mice and then contributed to the exacerbation of DSS-induced colitis, thereby proving the potentially crucial role of

Akkermansia muciniphila in gut homeostasis (Liu et al., 2022). Previous studies also suggested that treatment of *Akkermansia muciniphila* could ameliorate colitis symptoms by enhancing gut barrier integrity, decreasing inflammatory factors, and improving microbiota ecology (Bian et al., 2019). However, adverse effects of *Akkermansia muciniphila* in IBD have also been reported based on animal models. Intake of dietary sugars (such as glucose) could induce severe colonic inflammation in DSS-treated or *IL10*^{-/-} mice by modulating gut microbiota composition, in particular, the increase of *Akkermansia muciniphila* abundance could cause the decrease of the mucus layer and facilitate gut barrier damage during inflammation (Khan et al., 2020). Similar harmful effects

of *Akkermansia muciniphila* on IBD were also reported in *IL-10* knockout mice (Seregin et al., 2017). Those conflicting results and effects of gut microbiota on IBD could be explained by various factors, including the different genotypes of animal models and gut microbiota baseline. Another question is whether gut microbiota dysbiosis is a primary or secondary trigger for IBD. Numerous studies have emphasized the important roles of gut microbiota on IBD, however, recent evidence suggested that even inflammation *per se* could cause gut microbiota dysbiosis (de Souza and Fiocchi, 2016; Lupp et al., 2007). DSS treatment could induce colitis and gut dysbiosis in mice, however, DSS could not induce any significant changes in murine microbiota *in vitro*, including composition and metabolic pathways (Krause et al., 2024). Individual delivery methods can significantly influence the gut microbiota composition, but it is unlikely to substantially influence the incidence of IBD (Bruce et al., 2014). Those results demonstrated that gut microbiota dysbiosis alone may not be sufficient to trigger IBD. In this regard, more information about the interaction between gut microbiota and the host is desired.

2.2. Intestinal barrier

The first line of defense in human GI tracts is the intestinal barrier which can prevent the infiltration of potentially harmful compounds from intestinal content. In healthy individuals, the presence of an intact gut barrier, including epithelial cells and mucus layer, can prevent most bacteria or antigens from contact with lamina propria. The tolerance mechanism of humans can limit the dysregulation of immune cells and related inflammatory responses in the intestinal part. However, when the breach of the gut barrier happens, severe infiltration of luminal antigens can induce the failure of immune system tolerance and trigger the incidence of IBD (Kobayashi et al., 2020). Therefore, the gut barrier has attracted the attention of the academic community recently, and current evidence also proves that maintaining the gut barrier might be a crucial therapeutic target.

The mucus layer is mainly comprised of gel-forming mucins produced by goblet cells, acting as a crucial physical barrier at the top of epithelial cells. The structure of the mucus layer is different in the small intestine and colon. The small intestinal tract constitutes a single and penetrable layer, but in the colon, there are two layers of mucus, including a penetrable outer layer and an impenetrable inner layer (Johansson et al., 2013; Johansson and Hansson, 2016). The importance of the mucus layer in gut health has attracted the attention of researchers for many years, and the damage and defect of the mucus layer have been observed with decreased mucus layer thickness due to the decrease of mucin 2 expression (Van Klinken et al., 1999). The conduction of *Muc2*^{-/-} mice also illustrates the important role of mucus as a physical barrier in preventing luminal components infiltration and IBD (Van der Sluis et al., 2006). The relationship between gut microbiota and mucus is reciprocal, as previous experiment results also proved that the depletion of gut microbiota in germ-free and antibiotics-treated mice could reduce the thickness of the mucus layer in the colon via decreasing the expression of *Muc2* in the proximal colon part (Bergstrom et al., 2020). Furthermore, the mucus layer can also provide nutrients and habitat for gut microbiota, a prior study identified a distinctive gut microbiota community in *Muc2*^{-/-} mice which could further exacerbate DSS-induced colitis in wild-type mice after fecal microbiota transfer (Leon-Coria et al., 2021).

Immediately below the mucus barrier are the epithelial cells which maintain a high renewal rate and also act as a physical barrier to restrict the free passage of antigens from the intestinal lu-

men. The defect of gut epithelial cells has been linked to the trigger of IBD, for example, in the early stage of ulcerative colitis, the apoptotic foci could be found while the epithelium was normal under endoscopy test (Gitter et al., 2001). The GI tract is the most important digestion and absorption organ, thus, in addition to the crucial barrier functions, intestinal epithelial cells should also selectively allow the permeability of some nutrients and water. Tight junction proteins, including occludin, claudins, and zonula occludens (ZO)s, are the most crucial components for intracellular interactions between epithelial cells, equipping epithelial cells with those functions that seal the paracellular space and selectively restrict the entry of components (Chelakkot et al., 2018b). Due to the remarkable turnover rate of epithelial cells, the accurate regulation of tight junction proteins is required to maintain gut homeostasis. Current studies support that abnormal alterations of tight junction proteins can be detrimental to gut health and have been regarded as the crucial causes for the increase of gut permeability (Edelblum and Turner, 2009). For example, the decrease in occludin expression has been reported in active Crohn's disease (Zeissig et al., 2007), and the deficiency of *mdr1a* could trigger the development of spontaneous colitis in mice by impairing phosphorylation of tight junction proteins (Martini et al., 2017; Resta-Lenert et al., 2005).

Apart from forming a gut barrier, intestinal epithelial cells also play a crucial role as the interface between the immune regulation of the host and the gut microbiota. Some pattern recognition receptors have been found in intestinal epithelial cells, such as the Toll-like and NOD-like receptors, which can be constantly stimulated by gut microbiota-related molecular patterns and then activate epithelial cells to express antimicrobial peptides (including angiogenin-4) and cytokines (IL-33 and IL-25) (Hooper et al., 2003; Schiering et al., 2014; Zaph et al., 2008). Those factors are required for the forming of tolerogenic immune regulation which helps to preserve a symbiotic environment and regulate the gut microbiota ecology. Previous studies have revealed that the dysregulation of pathogen recognition receptor expression could be widely observed in patients with IBD, for example, the mutation of NOD-2 has been identified in patients suffering from Crohn's disease (Ogura et al., 2001). NOD-2 expression in intestinal epithelial cells has been linked with gut inflammation and homeostasis (Ferand et al., 2019). *Nod2*^{-/-} mice were also conducted to prove the functions of NOD-2 in the intestine, indicating that deficiency of NOD-2 could induce the expansion of *Bacteroides vulgatus* and exacerbate mucosal inflammation (Ramanan et al., 2014). Another example of the connections between gut microbiota, epithelium, and immune systems is the Toll-like receptor 4 (TLR4), which has also been connected with IBD and expressed in epithelial cells (Dheer et al., 2016). *TLR4*^{-/-} mice experienced enhanced susceptibility to colon inflammation due to the dysregulation of *Akkermansia muciniphila*-associated immune response (Liu et al., 2022).

2.3. Immune systems

The intestinal immune systems can be divided into innate and adaptive immunity protecting the gastrointestinal tracts from various diseases. In homeostasis conditions, the immune system maintains a delicate balance between numerous immune cells and contributes to the tolerance of humans to dietary factors and gut microbiota. However, the unbalanced modulation of immune systems has also been regarded as a potential trigger for the incidence of IBD.

The innate immune system, including neutrophils, macrophages, and dendritic cells (DCs), constitutes the first line of defense against any infiltration and exposure. The infiltration of neutrophils in intestinal tissues which can be observed throughout the

progress of active IBD is one of the earliest markers for gut-related inflammatory diseases (de Souza and Fiocchi, 2016). Neutrophils contribute to the progress of IBD via numerous mechanisms, such as neutrophil extracellular traps and myeloperoxidase which are also activated and upregulated in patients suffering from ulcerative colitis (Bennike et al., 2015; Dinallo et al., 2019). A prior study has also observed the crucial role of neutrophils in IBD with the conduction of *BATF3* deficiency mice, supporting that intestinal epithelium-derived *BATF3* in mice could promote colitis and even colonic cancer via increasing the number of neutrophils (Lin et al., 2021). The functions of macrophages in IBD are plastic, and macrophages have been classified as M1 and M2 with different cytokine patterns and various biological effects (Lissner et al., 2015; Wynn et al., 2013). M1-type macrophages perform important roles in promoting inflammation conditions via secreting cytokines, including IL-1 β , IL-6, TNF- α , etc., and promoting expression of inducible nitric oxide synthase (iNOS) (Yip et al., 2021). With the expression of those factors, macrophages can further influence the regulation of adaptive immune systems and promote IBD. In contrast, M2 macrophages have been classified as the anti-inflammatory type that can produce immunosuppressant factors, such as IL-10 and TGF- β (Yang et al., 2022). Previous research based on DSS-induced colitis in mice also indicated that colonic inflammation could be promoted via facilitating macrophage M1 polarization (Wei et al., 2020). Moreover, there are also some strategies tried to manipulate the polarization of macrophages to alleviate IBD (Liu, Ren et al., 2022). Besides secreting various cytokines, macrophages can also act as antigen-presenting cells to link innate and adaptive systems, and this similar function can also be observed in DCs, which can monitor the surrounding conditions and present antigens to induce adaptive immune responses. Intestinal DCs normally maintain a “hyporesponsive and tolerogenic state” in humans, however, studies implied an improper conditioning of DCs that were activated with high levels of TLR2 and TLR4 in patients suffering from IBD (Hart et al., 2005). Those dysregulated DCs could migrate to peripheral lymphoid tissues and further activate the antigen-specific response of T cells. The crucial role of DCs in the progress of IBD was also proved by previous studies that the DCs-derived CD134L was involved in the incidence of T cell transfer-induced colitis in murine models (Malmström et al., 2001).

In contrast to the innate immune systems, initiating adaptive immunity requires antigens and activated antigen-presenting cells. The gut homeostasis requires the delicate regulation between T cells, the key players of adaptive immunity. Previously, Crohn’s disease was postulated to be a Th1-driven condition due to the increasing expression of IL-12 and IFN- γ , while ulcerative colitis was closely linked with unconventional regulation of Th2 accompanied by elevated expression of IL-5 and IL-13 (Camoglio et al., 1998; Kobayashi et al., 2020). Th1 cells are crucial for eradicating intracellular pathogens, such as bacteria and viruses (Raphael et al., 2015), these cells can be induced with IL-12 produced by antigen-presenting cells upon antigens recognition (Gomez-Bris et al., 2023). In homeostasis condition, only a few CD4⁺ T cells could be observed in the gut, however, Th1 cells accumulated in the GI tracts of some patients with IBD and have been linked with the progress of diseases (Imam et al., 2018). IFN- γ has been applied as the defining cytokine expressed by Th1 cells, Nava et al. (Nava et al., 2010) demonstrated that IFN- γ could regulate intestinal epithelial homeostasis and loss of IFN- γ decreased colitis in mice models, indicating the potential pro-inflammatory of IFN- γ in IBD. However, some studies also mentioned the controversial role of IFN- γ in intestinal inflammation, Muzaki et al. (Muzaki et al., 2016) found that intestinal CD103⁺CD11b⁻ DCs improved IBD via IFN- γ -induced anti-inflammatory responses. Th1 cells

have also been defined as a source for the expression of TNF- α which has also been connected with the dysregulation of the intestinal barrier in intestinal inflammation and anti-TNF therapies have been applied for IBD treatment in patients (Kobayashi et al., 2020; Nava et al., 2010). Th2 cells normally contribute to the elimination of the parasite and participate in the regulation of immunity with other cells by producing various cytokines, including IL-4, IL-5, and IL-13 (Zeng, 2013), which can prevent the development of Th1 cells and activate the regulation of macrophages (Gomez-Bris et al., 2023; Raphael et al., 2015).

Besides the participation of Th1 and Th2 cells in IBD, emerging studies also demonstrated that there are some other subsets of CD4⁺ T cells contributing to the progress of IBD. Th17 cells have been regarded as pathogenic cells concerning the incidence of IBD, and previous clinical results observed an increased number of Th17 cells in patients suffering from IBD compared with healthy individuals (Annunziato et al., 2007; Caprioli et al., 2008; Lu et al., 2022). These cells can contribute to the regulation of immune systems via producing various cytokines, including IL-17, IL-21, and IL-22. Alexander et al. have demonstrated that gut microbiota can directly worsen DSS-induced colitis in mice via inducing the activation of Th17 cells and the expression of IL-17 (Alexander et al., 2022). Treg cells are essential for maintaining immunity tolerance, and the defect in Treg cells underlie autoimmune-related diseases and IBD (Mayne and Williams, 2013). Treg cells (CD4⁺FoxP3⁺CD25⁺) have been proven to be the existence in the mucosal of mice (Mottet et al., 2003). The crucial functions of these cells in IBD have also been identified with adoptive transfer colitis in mice, which induced colonic inflammation via adoptive transferring CD4⁺ T cells (CD4⁺CD45RB^{high} T cells) into *Rag1*^{-/-} mice (Kiesler et al., 2015). The initial IBD cause in this model has been linked to the lack of Treg cells in those transferred naïve CD4⁺ T cells. The anti-inflammatory and negative immunoregulatory roles of Treg in the gastrointestinal tracts can be exerted by numerous mechanisms, such as the expression of IL-10 and TGF- β (Vignali et al., 2008). Previous studies have highlighted the immunomodulatory functions of IL-10 with the conduction of *IL10*^{-/-} mice as spontaneously developed colitis models, showing the important role of IL-10 in the regulation of immune homeostasis (Gunasekera et al., 2020; Wilson et al., 2011).

Overall, these various and complicated mechanisms for the incidence of IBD present great challenges in understanding the interactions between lipid compounds and gut health. Dietary and gut microbiota-derived lipids may influence intestinal homeostasis and IBD progress through the mentioned one or several aspects.

3. Regulation of IBD by dietary lipids

The GI tract can digest and absorb nearly 95% of dietary lipid substances and those left lipids can therefore interact with gut microbiota. Accumulating results have suggested that dietary oils and fat could contribute to the modulation of gut microbiota composition and regulate gut homeostasis conditions (Murphy et al., 2015; Röytiö et al., 2017). With these studies, it has been shown that the amount of daily intake of different fats with varying fatty acid composition and other fat-soluble components can be linked with gut inflammation and homeostasis regulation.

3.1. High-fat diet

Previous studies have connected high-fat diets (HFD), especially

saturated fats, with systemic inflammation and diseases (Murphy et al., 2015). The gut microbiota as another fat consumer in humans can also be affected by HFD. Due to the exposure to dietary factors, the composition of these microbes colonizing in the gut also differentially develops reflecting the fluctuating diet. It has been demonstrated that HFD can induce a decrease in the richness and diversity of gut microbiota composition. A study has reported that HFD could induce a decrease in *Bacteroidetes* accompanied by an increase in *Proteobacteria* (Hildebrandt et al., 2009; Wolters et al., 2019). Although the clear mechanisms remain unknown, emerging studies have suggested that HFD-induced gut microbiota dysbiosis could act as a regulator to promote the related symptoms of IBD, demonstrating a promoted bacteria-derived endotoxemia as a direct evidence of increased gut permeability and increased lipopolysaccharide (LPS) in animal models and human (Cani et al., 2008; de la Serre et al., 2010; Serino et al., 2011). Moreover, research on HFD-induced gut dysbiosis has provided information for underlying mechanisms revealing the harmful effects of HFD-modulated gut microbiota on the intestinal barrier. HFD could reduce the abundance of some intestinal barrier-improving bacteria, including *Bifidobacterium*, *Lactobacillus*, and *Akkermansia muciniphilia*, accompanied by increasing bacteria lined with damaged barrier functions, including *Oscillibacter* and *Escherichia coli* (Anderson et al., 2010; Cani et al., 2007; Devkota et al., 2012; Lam et al., 2015; Lin et al., 2016; Sun et al., 2016; Sun et al., 2020). Although the clear mechanisms of the interactions between those specific bacteria and the gut barrier are still unclear, bacteria-derived metabolites may play a role in the maintenance of the gut barrier. Extracellular vesicles produced by *Akkermansia muciniphilia* play a crucial role in controlling gut barrier functions by promoting the expression of tight junction proteins (Chelakkot et al., 2018a). Similarly, *Bifidobacterium* and *Lactobacillus* may have beneficial effects on improving gut barrier functions and tight junction proteins via unknown mechanisms (Anderson et al., 2010; Hsieh et al., 2015). In contrast, the HFD-induced increased bacteria, *Oscillibacter*, has been directly linked to a decrease in the expression of tight junction proteins and an improvement of intestinal barrier (Lam et al., 2012). At this point, an increase in LPS associated with HFD feeding has also been found to modulate tight junction proteins and promote permeability in cell levels by activating the NF- κ B pathway (Guo et al., 2013). Moreover, *Escherichia coli* as the pathogenic microorganism is also able to modulate the barrier functions by degrading the mucins and damaging the mucus layer with the expression of proteases (Paone and Cani, 2020). In addition, previous studies have also emphasized that the gut bacteria from HFD-treated mice was able to activate some pro-inflammatory pathways in germ-free mice, proving the potential interactions between HFD-modulated gut microbiota and immune systems (Ding et al., 2010). The pro-inflammatory effects of HFD on the intestinal tracts are normally linked with the increase of LPS which is a pro-inflammatory factor in immune systems, for example, LPS can induce the polarization of M1 macrophages (Zheng et al., 2013). Meanwhile, the decrease in some specific bacteria may also induce the modulation of immune systems. With the conduction of *TLR4*^{-/-} mice, Liu et al. reported that the decrease of *Akkermansia muciniphilia* might induce the dysregulation of ROR γ ^t Treg cell and further exacerbated DSS-induced colitis in mice (Liu et al., 2022). Interestingly, the influence of HFD on gut microbiota can also modulate the gut homeostasis and development of offspring. Xie et al. (Xie et al., 2018) found that the consumption of HFD during pregnancy could induce gut microbiota dysbiosis and low-grade inflammation in offspring, even further promoting susceptibility to experimental colonic inflammation in adulthood.

In addition to the gut microbiota that has been described, HFD can also modulate gut homeostasis and contribute to the progress of IBD via direct or indirect ways. Recent studies have pointed out that the potential influence of HFD on the composition and functions of the mucus layer is mainly to affect its major protein, mucin, which is a high molecular weight and heavily O-glycosylated protein expressed by goblet cells. For example, HFD feeding for 25 weeks changed the oligosaccharide chain of mucin in mice and further contributed to colon inflammation (Mastrodonato et al., 2014). Although clear mechanisms were not provided in this study, the influence of HFD feeding on mucin was hypothesized to be connected with the glycosylation defect or incomplete maturation of goblet cells. The consumption of HFD can also induce endoplasmic reticulum /oxidative stress in goblet cells, triggering the unfolded protein response and damaging the formation of mucus layer (Gulhane et al., 2016). Moreover, in the small intestine, HFD can modify the PPAR- γ pathway in mice to repress the expression of *Cftr* gene, leading to a reduction in chloride and loss of mucus barrier integrity, which may contribute to the incidence of IBD (Tomas et al., 2016). Other studies also parsed out the influence of HFD on other components of the intestinal barrier, for example, an HFD pattern reduced the expression of tight junction proteins and thus promoted gut permeability in mice (Cani et al., 2008; Kirpich et al., 2012). Similarly, effects of HFD on tight junction proteins were also observed in obese (Otsuka Long Evans Tokushima Fatty) and lean rat (Long Evans Tokushima Otsuka) strains, indicating that the suppression of tight junction proteins could be mainly attributed to HFD rather than obesity and related modulated metabolism (Suzuki and Hara, 2010). Apart from the influence of HFD-modulated gut microbiota, the direct impact of HFD on tight junction proteins was also observed based on related cell models. Directly treating Caco-2 cells with dietary fatty acids contained in HFD could induce promoted tight junction permeability via protein kinase C (PKC) regulation (Usami et al., 2003). Those results indicated that HFD can also interact with intestinal epithelium and change its functions. With these concepts, other studies also found that the HFD could also induce gut barrier dysfunction by increasing the apoptotic rate of intestinal epithelial cells. The accumulation of long-chain fatty acids in the intestine with HFD treatment has been proven as a potential cause to increase ROS production, mitochondrial dysregulation, and pro-activating apoptotic-related pathways (Li and Li, 2020). Moreover, the consumption of HFD can also modulate the bile acids profile, such as the increase of taurocholic acid and deoxycholic acid, then inducing the damage of intestinal epithelial cells (Barrasa et al., 2011; Wan et al., 2020; Wolf et al., 2020).

HFD has also been connected with the disordered regulation of immune systems in IBD. In the DSS-induced colitis model, the treatment of HFD exacerbated the symptoms of inflammation accompanied by decreased Treg cells in the colon (Ma et al., 2007). Apart from the influence of HFD-changed gut microbiota and bile salts on the intestine (Zhao et al., 2020), there are emerging studies that have reported the potential effects of HFD-induced obesity and adipose tissue on immune dysregulation in IBD and related diseases. HFD-induced obesity could accelerate experimental colitis-associated colorectal cancer by exacerbating inflammation and dysregulation of immune systems (Wunderlich et al., 2018). The obesity-exacerbated colonic cancer has been mainly attributed to the shifted macrophage polarization by obesity-induced IL-6, which further expresses CCL-20 and recruits CCR-6-expressing B cells and $\gamma\delta$ T cells. Similarly, Wei et al. also revealed that HFD could modulate the miRNA profiles of visceral adipose exosomes, changing these exosomes from anti-inflammation to pro-inflammation type. Interestingly, those pro-inflammatory exosomes can

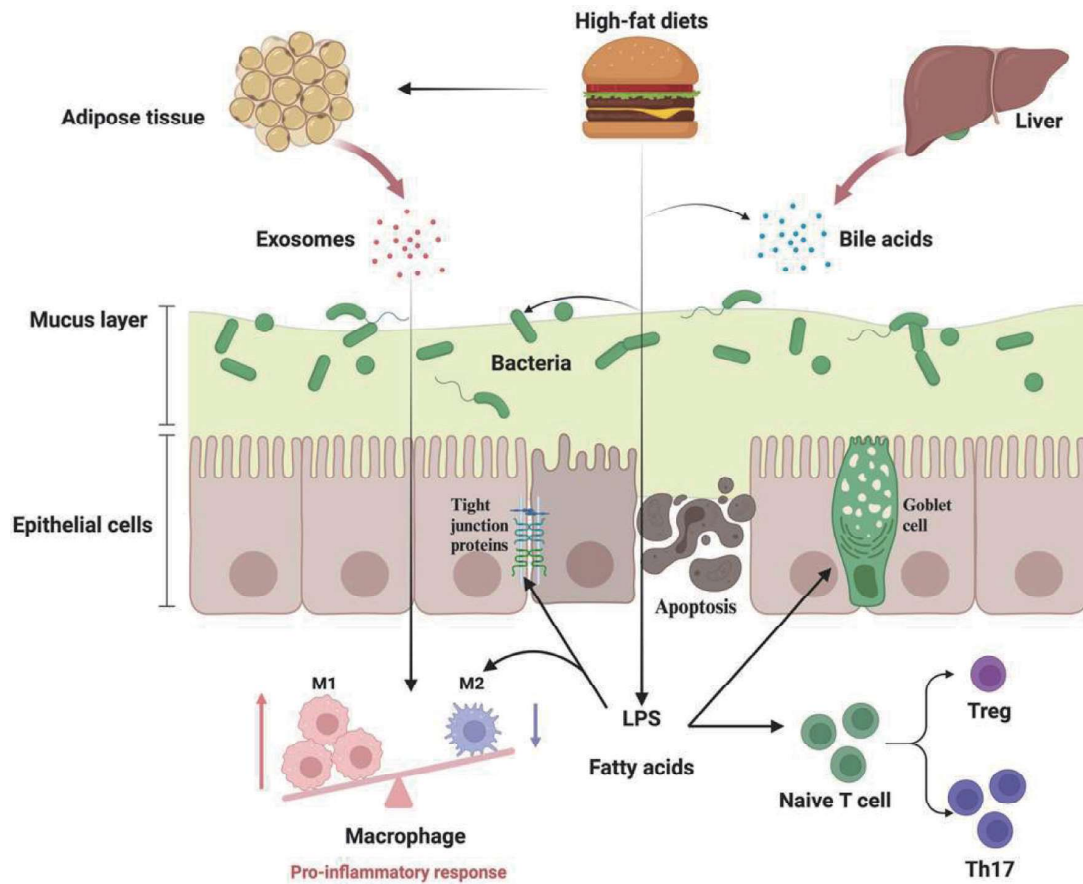


Figure 2. The potential effects of high fat diets (HFD) on intestinal homeostasis and inflammatory bowel disease (IBD). The HFD-induced dysbiosis of gut microbiota, including the decrease in richness and imbalance of the *Firmicutes/Bacteroidetes* ratio, has been widely linked with bacteria-derived lipopolysaccharide (LPS) that can further trigger low-grade inflammation and immune dysregulations. The mucus layer, a crucial physical barrier in the intestinal, can also be damaged with HFD due to the dysfunction of goblet cells and alteration of peroxisome proliferator-activated receptor gamma (PPAR- γ) pathway. HFD can also induce gut permeability by modulating tight junction proteins and epithelial cell apoptosis. HFD-induced obesity is also associated with the progress of IBD via promoting M1 macrophage differentiation and recruiting lymphocytes. Figure 2 was created in [Biorender.com](#) (Agreement number: MC27CARFOD).

then circulate to the GI tracts and exacerbate DSS-induced colitis in mice by enhancing M1 macrophage differentiation (Wei et al., 2020) (Figure 2).

3.2. Ketogenic diet

A ketogenic diet with about 90% calories from fat has been practiced by many people for weight loss and blood sugar control (Hall and Chung, 2018). Ketogenic diets are effective in achieving ketosis which is an anti-inflammatory state in humans with an increase in blood acetoacetate and β -hydroxybutyrate. Even though it is controversial, some studies also found that ketogenic diets could affect the gut microbiota composition in humans with a promotion abundance of some SCFAs-producing bacteria (such as *Lactobacillus* and *Bacteroides*) (Alsharairi, 2022). Due to the nonnegligible roles of SCFAs in IBD, it was also hypothesized that the consumption of the ketogenic diet might exert beneficial effects on IBD patients. The significant impact of ketogenic diets on gut microbiota composition was also detected in another inpatient crossover study (Ang et al., 2020).

The effects of ketogenic diets on IBD and the potential under-

lying mechanisms have been examined based on animal models. The modulation effects of ketogenic diets on gut microbiota were also observed in mice, accompanied by a significant decrease in *bifidobacteria* (Ang et al., 2020). Furthermore, this study showed that the adjusted gut microbiota composition in ketogenic diet-fed mice could be mainly attributed to the increased level of ketone bodies. As a consequence of ketogenic diet-induced gut microbiota alteration, this dietary pattern could reduce the levels of Th17 cells in the intestinal tissues, emphasizing the potential anti-inflammatory effects of ketogenic diets in IBD. Analogical effects of ketogenic diets on gut microbiota and immune systems were also observed in mice with DSS-induced colitis, Kong et al. suggested that ketogenic diets could alleviate colonic inflammation by decreasing the production of ROR γ t⁺CD3⁻ group 3 innate lymphoid cells (Kong et al., 2021). With the conduction of germ-free mice and FMT experiments, the beneficial effects of ketogenic diets on colitis and immune system regulation have been connected with the regulation of gut microbiota composition. Conversely, another study also demonstrated that ketogenic diets could aggravate DSS-induced colitis in mice with promoted inflammatory symptoms and gut microbiota dysbiosis when compared with control diet-fed

Table 1. The main effects of dietary fatty acids on the gut microbiota

Dietary fatty acids	Type	Model	Effect on gut microbiota	References
Saturated fatty acids (SFAs)	lard oil	Mice fed high fat diet	diversity and richness decreased	Liu et al., 2019
	milk-derived SFA	<i>IL10</i> ^{-/-} mice	<i>Bilophila wadsworthia</i> increased	Devkota et al., 2012
	N.A	Human	proinflammatory bacteria like <i>Proteobacteria</i> increased	Mandal et al., 2016
Medium-chain fatty acids (MCFAs)	glycerol monolaurate	Colitis Mice	<i>Lactobacillus</i> and <i>Bifidobacterium</i> increase	Mo et al., 2021
	N.A	Mice fed high fat diet	decreased <i>Firmicutes/Bacteroidetes</i> ratio and <i>Proteobacteria</i>	Zhou et al., 2017
Monounsaturated fatty acids (MUFAs)	palmitoleic acid	Colitis Mice	<i>Akkermansia muciniphila</i> increased	Chen et al., 2023b
	oleic acid	Colitis Mice	anti-inflammatory bacteria increased	Fernández et al., 2020
	oleic acid	Mice fed high fat diet	total bacteria density increased	Mujico et al., 2013
Polyunsaturated fatty acids (PUFAs)	ω-3 fatty acids	Human	<i>Lachnospiraceae</i> increased	Menni et al., 2017
	ω-3 fatty acids	Colitis Mice	<i>Akkermansia</i> , <i>Alistipes</i> , and <i>Lactobacillus</i> increased	Dong et al., 2022
	conjugated linoleic acid	Colitis Mice	<i>Bacteroides</i> decreased, <i>Bifidobacterium</i> increased	Chen et al., 2019
	linoleic acid	Colitis Mice	<i>Escherichia coli</i> increased	Deol et al., 2023

N.A means not available.

groups (Li et al., 2021). Some human studies and investigations also mention the potential negative influence of ketogenic diets on gut health and microbiota, highlighting the necessity to further examine the safety and clear mechanisms of ketogenic diets and gut ecology (Gentile and Weir, 2018).

3.3. Fatty acids

In addition to the amount of total dietary fat, the composition of fatty acids could also influence the gut microbiota and even induce harmful or beneficial effects (Table 1). Previous studies have suggested that the composition of gut microbiota is significantly different between murine models treated with polyunsaturated fatty acids enriched fish oil and saturated fatty acids enriched with lard (Li et al., 2017). Meanwhile, Haskey et al. (2022) compared the influence of different dietary oils on the progress of IBD by conducting *Muc2*^{-/-} mice as a model, implying that the fatty acid composition of fat might also impact colitis in mice.

3.3.1. Saturated fatty acids (SFAs)

Numerous studies have demonstrated that SFAs are potential pro-inflammatory dietary factors for human health, especially in gastrointestinal tracts. The intake of lard oil could significantly dysregulate the composition of gut microbiota with a decrease in diversity and richness (Liu et al., 2019). The previous results also reported the potential connections between SFAs-induced gut microbiota dysbiosis and IBD, Devkota et al. (2012) showed that the treatment of SFAs (milk-derived) could promote the expansion of *Bilophila wadsworthia* which is a kind of pathobiont and exacerbate colitis in *IL10*^{-/-} mice with Th1 immune response. Some clinical studies also emphasized the potentially harmful influence

of dietary SFAs on gut microbiota in humans, for example, the increase in SFA consumption has been linked to the modulation of some pro-inflammatory bacteria (including *Proteobacteria*) during pregnancy (Mandal et al., 2016). These studies clearly revealed that the intake of SFAs could modulate the composition of gut microbiota, and this disturbed bacterial ecology may further aggravate intestinal inflammation.

Although the mechanisms remain unclear, the direct pro-inflammatory effects of SFAs in the gastrointestinal tract have also been reported in previous studies. It has been revealed that SFAs might increase intestinal permeability and serum LPS amount by damaging the gut barrier. Ghezal et al. (2020) showed that palmitic acid treatment could decrease the expression of tight junction proteins and cause their mislocalization between cells, leading to a decrease in the mucosal barrier. This study also demonstrated that the SFA treatment could promote the expression of pro-inflammatory factors in mice, including IL-1β. The similar pro-inflammatory effects of SFAs-contained diets have been widely proven, the administration of myristate and stearate could induce proinflammation activation via activating IL-1β and TNF-α in human monocytes, whereas these effects could not be induced with the unsaturated fatty acids (Pillon et al., 2016). The pro-inflammatory pathway, NF-κB, could also be modulated with the treatment of SFAs (Lee et al., 2003). In summary, these studies clearly showed that SFAs and related diet patterns may negatively influence gut permeability and these dietary factors can also promote the production of inflammatory-related factors, which may further contribute to the progress of IBD.

3.3.2. Medium-chain fatty acids (MCFAs)

MCFAs are normally characterized as fatty acids with shorter carbon chains (C6-C12) that can be directly absorbed by humans and

quickly supply energy (Jia et al., 2020a). Nowadays, the potential influence of MCFAs on IBD has been elucidated with some clinical and epidemiological results. With the analysis of fecal samples with the GC-mass method, significantly decreased MCFAs (including pentanoate, hexanoate, heptanoate, and octanoate) have been reported in IBD patients, and MCFAs have been regarded as a potential metabolic biomarker (De Preter et al., 2015). Similar results were also shown in adolescent patients with ulcerative colitis, the amount of lauric acid in the blood was higher during the active phase compared with quiescent conditions (Kikut et al., 2022). These results supported that MCFAs might play a crucial role in the treatment and prediction of IBD. However, the actual application of MCFAs as the medical therapeutic for IBD is still challenged, while the supplementation of MCFAs may cause the deficiency of other functional fatty acids and fat-soluble nutrients (Łoś-Rycharska et al., 2016).

MCFAs can improve the progression of IBD by modulating inflammatory factors and the gut barrier. The treatment of lauric acid could decrease the promoted level of some inflammatory factors via modulating TLR4/MyD88 pathway in rats with LPS-induced inflammations (Khan et al., 2021). Moreover, glycerol monolaurate, the mono-ester formed from glycerol and lauric acid, was reported that can attenuate DSS-induced colitis by promoting colonic Foxp3⁺ Tregs cells and ratio of serum anti-inflammatory/proinflammatory cytokines, as well as reconstructing microbial communities (Mo et al., 2021). The Black Soldier Fly Larvae's oil which contains a high amount of MCFAs could ameliorate colitis in mice (Richter et al., 2023). Further results from transcriptome analysis provided potential mechanisms that these functional oil components could regulate mTOR signaling and promote the expression of PPAR-related genes. The beneficial effects of MCFAs were also observed in TNBS-induced colitis in mice, while the treatment of dietary medium-chain triglycerides could alleviate colitis with the inhibition of IL-1 β , TNF- α , and MPO expression (Kono et al., 2010). The impact of MCFAs on the gut barrier has also been examined in epithelial cell lines, caprylic and nonanoic acid could reduce bacterial translocation and enhance barrier functions by attenuating the activity of the histone deacetylase pathway (Wang et al., 2018). Based on the samples from IBD patients and healthy individuals, the influence of sodium caprate on paracellular permeability and tight junction proteins has also been confirmed. Although clear information is still limited in IBD, the gut microbiota regulation roles of MCFAs have also been reported. MCFAs could prevent body weight gain in high-fat diet-treated mice and modulate the gut microbiota composition, including decreased *Firmicutes/Bacteroidetes* ratio and *Proteobacteria* (Zhou et al., 2017). Moreover, an increase in SCFAs was also induced with the treatment of MCFAs, emphasizing the potential beneficial role of MCFAs on gut ecology.

3.3.3. Monounsaturated fatty acids (MUFAs)

MUFAs that contain a carbon double-bond and mainly include palmitoleic and oleic acid can be found in the human daily diets, such as olive oil and nuts (Las Heras et al., 2022). Numerous studies have linked MUFAs with some beneficial effects on gut microbiota and inflammation-related diseases (Statovci et al., 2017). Among MUFAs, the effective anti-inflammatory activities of palmitoleic acid in IBD have been reported. When cultured colonic tissue of IBD patients was treated with palmitoleic acid, the expression of inflammation-related factors, such as IL-6 and TNF- α , was significantly reduced (Chen et al., 2023b). This study also observed that the oral intake of palmitoleic acid could allevi-

ate experimental colitis in mice models and reshape gut microbiota composition with an increase in the abundance of *Akkermansia muciniphila*. Macrophage is another potential target, chronic palmitoleic acid supplementation can reduce the systemic level of IL-1 β by evoking the lipidomic remodeling of the endoplasmic reticulum in macrophages (Çimen et al., 2016). Souza et al. also reported that palmitoleic acid could decrease LPS-induced pro-inflammatory responses in macrophages and alter the ratio of M1/M2 in a PPAR α -independent manner (Souza et al., 2017).

The effects of another MUFA (oleic acid) on gut health and related mechanisms are unclear. With the application of 7-day food diaries in prospective cohort investigation, oleic acid was recognized as a beneficial dietary factor in decreasing the incidence of ulcerative colitis (de Silva et al., 2014). The animal studies based on rats treated with acorn-fed ham which contains high levels of oleic acid supported the potential intervention role of oleic acid in the management of IBD (Fernández et al., 2020). In this study, the consumption of oleic acid could alleviate the symptoms of IBD and also reduce the amount of some proinflammatory factors (including IL-17 and IFN- γ). The reconstruction of gut microbiota composition was also mentioned in this model, showing that the treatment of oleic acid-enriched diets was sufficient to promote the abundance of some anti-inflammatory bacteria and the concentration of SCFAs. Similarly, supplementation of oleic acid could significantly counteract the HFD-perturbed gut microbiota dysregulation, accompanied by increased total bacteria density (Mujico et al., 2013). However, some contradictory results were also reported, a clinical study previously compared the influence of two dietary patterns with different lipid compositions (79% and 28% oleic acid) on patients suffering from active Crohn's disease, reporting that the remission rate of the higher oleic acid treated group was lower (Gassull et al., 2002). Furthermore, some studies also supported that the intake of oleic acid might promote the risk of the incidence of IBD (Sugihara et al., 2018; Ye et al., 2021). Therefore, more studies and clinical data are still required to further clarify the influence of oleic acid on gut health and diseases based on various models.

3.3.4. Polyunsaturated fatty acids (PUFAs)

PUFAs, including ω -3 and ω -6 fatty acids that contain multi-double bonds, are also abundant fatty acids in human diets. The supplement of PUFAs is crucial for many biological activities in humans because they cannot be directly produced by humans and daily diets are the major exogenous resource. Numerous studies have been conducted to declare the potential beneficial effects of ω -3 fatty acids including α -linolenic acid (ALA), eicosapentaenoic acid (EPA), and docosahexaenoic acid (DHA). However, there are still some debates about the influence of ω -6 fatty acids including linoleic acid (LA) and arachidonic acid (AA) as many studies conflict with others and generate the mixed results of the potential pro-inflammatory or anti-inflammatory activity.

Accumulating evidence based on clinical trials suggested the crucial roles of PUFAs in the progression of IBD. A decreased level of serum ω -3 fatty acids has been found in patients suffering from ulcerative colitis and Crohn's disease compared with normal individuals (Jiang et al., 2023). A previous 170,805 women included prospective study suggested that cumulative energy-adjusted consumption of PUFAs could not promote the incidence and risk of IBD, and the excessive dietary ω -3 fatty acids could be linked with a lower risk of ulcerative colitis (Ananthakrishnan et al., 2014). Similar anti-inflammatory effects of ω -3 fatty acids were also supported by a previous Mendelian randomization analysis,

demonstrating the beneficial role of ω -3 fatty acids in IBD (As-tore et al., 2022). Different from the desired influence of dietary ω -3 fatty acids in IBD, numerous studies have concluded that ω -6 fatty acids could be associated with increasing IBD pathogenesis. Dietary LA is metabolized to AA and then causes a direct influence on health. In patients with active IBD, the amount of AA was significantly increased compared with healthy individuals, and these results have been partially attributed to the promoted infiltration of inflammatory cells (Nishida et al., 1987). A prospective cohort study that involved 203,193 individuals was also conducted further to uncover the role of LA in IBD and demonstrated that increased LA consumption could contribute to the promoted risk of IBD (Tjonneland et al., 2009). Conversely with other ω -6 fatty acids, conjugated linoleic acid (CLA) could exert an anti-inflammatory activity in patients with Crohn's disease by suppressing the expression of IL-17, IFN- γ , and TNF- α from CD4⁺ and CD8⁺ T cells (Bassaganya-Riera et al., 2012).

Dietary PUFAs can influence the incidence and pathogenesis of IBD by directly modulating inflammatory-related factors and pathways. The clear role of ω -3 fatty acids in IBD was discovered based on transgenic *fat-1* mice that could produce ω -3 fatty acids without any diet supplement. Those endogenous ω -3 fatty acids could relieve DSS-induced colitis and the anti-inflammatory effects have been partially connected with the suppressed NF- κ B activity (Hudert et al., 2006). Similar results were also observed in rats with experimental-induced Crohn's disease, intake of ω -3 fatty acids could improve the symptoms of IBD accompanied by an increase in the activation of PPAR- γ (Yao et al., 2017). Although underlying mechanisms are still unclear, various studies have been performed to investigate the influence of some individual ω -3 fatty acids on IBD. For example, previous studies have shown that oral intake of ALA could improve DSS-induced colitis in mice (Kim et al., 2020). The anti-inflammatory effects of ALA were further revealed in Caco-2 cells, where the treatment of ALA could inhibit the expression of IL-8, COX2, and iNOS (Reifen et al., 2015a). The administration of EPA or DHA has also been supported as a potential measurement for IBD management based on animal results, showing that intake of EPA and DHA could attenuate DSS-induced colitis. This study also emphasized that EPA was more effective compared with DHA, and the underlying mechanisms might involve the NLRP3/IL-1 β and IL-6/STAT3 inflammation-related pathways (Zhang et al., 2021). The exacerbation effects and potential mechanisms of ω -6 fatty acids have also been discussed based on *in vivo* and *in vitro* models. A diet rich in LA could accelerate and exacerbate colitis in *IL10*^{-/-} and wild-type mice (Deol et al., 2023). The metabolomic analysis showed that these adverse effects of LA could be linked to the increase of oxylipins and the decrease in endocannabinoid system metabolites. In patients with Crohn's disease, the decreased activity of glutathione peroxidase 4 (GPX4) was observed in the small intestinal epithelial cells. The previous studies indicated that an AA diet could induce intestinal inflammation in *Gpx4*^{+/-}-IEC mice by triggering cytokine production similar to ferroptosis (Mayr et al., 2020). Different from LA and AA, intake of CLA could ameliorate chemically induced colitis in a dose-dependent manner, accompanied by decreasing inflammatory factors and improving gut barrier (Chen et al., 2019). Another study further provided evidence based on *in vivo* experiments and declared that the protective effects of CLA on IBD could be partly due to the activation of PPAR γ and δ (Bassaganya-Riera et al., 2004).

The dysbiosis of gut microbiota plays a crucial role in the progression of IBD, and PUFAs may impact IBD via modulating gut bacterial ecology. The associations between dietary ω -3 fatty acids and gut microbiota have been observed based on the data from 876 twins, emphasizing that ω -3 fatty acids supplementation could

improve microbiome composition, especially for *Lachnospiraceae* (Menni et al., 2017). In the colitis mice model, ω -3 fatty acids could also improve the inflammation symptoms and modulate the composition of gut microbiota, including some differential bacterial *Akkermansia*, *Alistipes*, and *Lactobacillus* (Dong et al., 2022). Moreover, the beneficial effects of CLA on IBD have also been partly attributed to its modulation functions on gut microbiota. The administration of 40 mg CLA per day could rebalance the DSS-induced dysbiosis in gut bacteria, while a decrease in *Bacteroides* and an increase in *Bifidobacterium* have been reported in the CLA-treated group (Chen et al., 2019). Promoting effects of LA on gut dysbiosis have also been revealed in previous study, which reported an increased abundance of *Escherichia coli* in LA-fed mice and this bacteria might contribute to the exacerbation of IBD (Deol et al., 2023). Conflicts in the interactions between PUFAs and gut microbiota should also be mentioned, the intake of high-dose omega-3 PUFAs could not directly induce significant modulation effects on gut microbiota in healthy humans (Watson et al., 2018). These results clarified that the influence of PUFAs on gut microbiota and IBD is still unclear, and the potential effects of PUFAs on bacterial ecology may also be affected by other molecules or the health conditions of individuals. Due to the complexity and importance of the gut microbiome, more detailed and well-designed experiments (such as germ-free mice and FMT) are desired to be conducted to provide more information.

3.4. Cholesterol

Cholesterol is present in our daily diets and humans intake around 200 to 600 mg per day (Liu et al., 2023). It performs numerous crucial roles in humans, including maintaining the fluidity and structure of the cell membrane and serving as the precursor for the production of sex hormones and vitamin D (Olkkonen et al., 2017). However, the excessive intake of dietary cholesterol can be associated with the high risk of cardiovascular diseases (Zhong et al., 2019) and even cancer (Hu et al., 2012). The daily diet is one of the major sources of cholesterol for humans, and this exogenous cholesterol can enter the small intestine as micelle forms. It has been reported that only 50% of cholesterol is absorbed in the small intestine, indicating that the direct interaction between dietary cholesterol and colon cannot be negligible (van der Wulp et al., 2013). Meanwhile, the absorbed cholesterol could be applied to synthesize bile acids in the liver and further released to the gastrointestinal tracts which may also have some impacts on the homeostasis conditions of the gut (Collins et al., 2023). With those direct and indirect interactions between dietary cholesterol and the gut, the potential influence of high-cholesterol diets on IBD has received some attention.

Prolonged consumption of high-cholesterol diets has been connected with various chronic and inflammatory diseases in intestine tracts. Previous studies have also demonstrated that intake of high-cholesterol diets could induce the acute inflammation response in the intestine with an IL-1 β -dependent increase of myeloid cells in both murine and zebrafish models (Progatzky et al., 2014). These inflammatory responses during the treatment of high-cholesterol diets have been mainly attributed to the direct influence of ingested dietary cholesterol on the inflammasome activation in epithelial cells. Concerning the potential influence of high-cholesterol diets on inflammasome, some studies have investigated the interactions between high-cholesterol diets and inflammasome-related diseases. Du et al. (Du et al., 2016) reported that the intake of high-cholesterol diets could promote inflammatory responses in the colon by the activation of NLRP3 inflammasome. Gao et al. (Gao et al., 2010) found that cholesterol diets could weaken the colon unfold-

ed protein responses and then exacerbate colitis in mice, causing gut barrier damage.

Dietary cholesterol may also modulate the gut microbiota composition. Previous studies have reported that the long-term intake of high-cholesterol diets could drive liver cancer by altering the composition and metabolites of gut microbiota in mice (Zhang et al., 2021). Hao et al. (Hao et al., 2019; Hao et al., 2020) have also reported that the consumption of high-cholesterol diets could modulate the composition of gut microbiota and cause a decrease in short-chain fatty acids (SCFAs), including acetic, propionic, and butyric acids. As a favorable modulator of the immune system, gut barrier, and gut microbiota, the decrease in SCFAs has been regarded as a potentially harmful factor for the incidence of IBD (Caetano and Castelucci, 2022). Therefore, the potentially harmful effects of high-cholesterol diets on IBD may also be partially mediated by gut microbiota.

3.5. Oxidized cholesterol (OXC)

Dietary cholesterol is susceptible to oxidation to produce oxidized cholesterol (OXC) during various food processing stages (Xu et al., 2011). In diets, the oxidation of cholesterol mainly occurs with reactive oxygen species, and numerous kinds of OXC have been found in human diets, especially 7 α -hydroxycholesterol, 7 β -hydroxycholesterol, 5,6 β -epoxycholesterol, 5,6 α -epoxycholesterol, and 7-ketocholesterol. OXC has been widely regarded as a potentially harmful substance for human health (Deng et al., 2023). Furthermore, the amount of OXC in Western diets that contain high concentrations of cholesterol cannot be ignored. Previous studies have shown that the intake of OXC might reach up to 10% of cholesterol (Xu et al., 2009; Xu et al., 2011). OXC may be more toxic and harmful compared with naïve cholesterol in diets (Maldonado-Pereira et al., 2018).

OXC may induce oxidative stress and exert pro-inflammatory effects in humans. OXC has also been shown to be a potential trigger for IBD. The amount of serum OXC was significantly higher than cholesterol in patients suffering from IBD, suggesting potential associations between OXC and IBD (Akerlund et al., 1994). In this regard, Bai et al. (Bai et al., 2005) reported that 25-hydroxycholesterol significantly promoted the expression of IL-8 with the treatment of IL-1 β in Caco-2 cell models. The pro-oxidant and pro-apoptosis effects of OXC mixture (including 7 α -hydroxycholesterol, 7 β -hydroxycholesterol, 7-ketocholesterol, 5 α ,6 α -epoxycholesterol, 5 β ,6 β -epoxycholesterol) were also observed at the cell level (Biasi et al., 2009). A previous proteomic study based on intestinal epithelial cells illustrated that the treatment of 7-ketocholesterol could affect the expression of proteins related to inflammation and mitochondrial functions (Laparra et al., 2015). Other studies also observed that OXC could stimulate the activation of caspase-3, enhance the expression of various inflammation-related factors, including TLR2, TLR9, and IL-6 (Biasi et al., 2009; Guina et al., 2015; Mascia et al., 2010). Exogenous OXC may also impair the function of the gut barrier and promote intestinal permeability. Chalubinski et al. (Chalubinski et al., 2014) found that the treatment of 7-ketocholesterol could damage the barrier function of the human intestinal epithelium, accompanied by a decrease in mRNA expression of ZO-1. OXC-induced fall of IL-10 expression was also observed in these studies, indicating that OXC might induce inappropriate inflammatory regulation. Similar results about the effects of OXC on the permeability of Caco-2 cells were also reported in other studies, finding that OXC could decrease levels of ZO-1, occludin, and junctional adhesion molecule-A (JAM-A) in OXC-treated cells (Deiana et al., 2017). Interestingly, the effects of OXC on intestinal barrier func-

tion were also associated with the induction of matrix metalloproteinase-2 and -9 (MMP-2 and MMP-9) which had been shown to cause gut barrier damage via the degradation of junctional proteins (Meijer et al., 2007; Nighot et al., 2015).

The underlying mechanisms of the adverse effects of dietary OXC on IBD have been hypothesized to be mediated by gut microbiota based on recent animal-related research. The administration of 27-hydroxycholesterol via subcutaneous injection could induce gut microbiota dysbiosis in mice with a decrease in *Roseburia* and SCFAs, indicating the potential influence of OXC on gut microbiota (Wang et al., 2020). Similar aggravated gut microbiota dysbiosis was also observed in mice models with DSS-induced colitis, showing that exposure to OXC could exacerbate chemically induced inflammation in the intestine tracts with significantly increased body weight loss, colon length decrease, and bleeding (Yan et al., 2022). In this study, the gut microbiota composition was regulated by OXC with an increase in some harmful bacteria, such as *Escherichia-Shigella* and *Bacteroides*. With the further conduction of antibiotic treatment and FMT experiments, the exact mechanisms of OXC-promoted colitis in mice have been further revealed, and the crucial role of OXC-modulated gut microbiota was also uncovered (Yan et al., 2023). However, due to the difference between the gut microbiota in humans and mice, the influence of dietary OXC on patients with IBD is still limited, and some clinical or epidemiological investigations are still desired.

3.6. Fat soluble vitamin

Some fat-soluble components in dietary fat also play some roles in metabolic regulation and physiological functions. Recently, the potential influence of dietary fat-soluble vitamins (including vitamins A, D, and E) on gut ecology and inflammation has been examined. It had been shown that fat-soluble vitamins were quantitatively lower in patients with IBD compared with healthy individuals, emphasizing that their appropriate supplementation may improve these inflammatory diseases (Fabisiak et al., 2017).

3.6.1. Vitamin A

The deficiency of vitamin A is a worldwide health issue, especially in some developing countries. A previous study has implied that the deficiency of vitamin A in children could increase the risks of respiratory and diarrheal infections, and these harmful effects could be simply reversed with the intake of adequate vitamin A (Villamor and Fawzi, 2000). Moreover, the composition and dysbiosis of gut microbiota in humans may also be influenced by vitamin A. Human subjects with persistent diarrhea were selected to evaluate the effects of vitamin A deficiency on gut microbiome, and results showed a decrease in bacterial diversity in the vitamin A-deficient group with an increase in *Enterococcus* (Lv et al., 2016). Considering the potential connections between vitamin A and the incidence of IBD, the efficiency of vitamin A treatment on IBD has also been carried out. A daily intake of vitamin A (25,000 IU) for two months could significantly improve the clinical symptoms of ulcerative colitis, emphasizing the potential positive effects of vitamin A in IBD (Masnadi Shirazi et al., 2018). Various experiments have also been conducted in animal models to further support the underlying mechanisms for vitamin A supplementation. The influence of vitamin A deficiency on gut microbiota composition and ecology was also observed in mice. Results showed a decrease in butyrate in the vitamin A deficient group (Cha et al., 2010; Tian et al., 2018). The analogical influence of vitamin A treatment on the regulation

of gut microbiota was also shown in DSS-induced mice. The oral intake of vitamin A could ameliorate colonic inflammation in mice with an increase in SCFAs-producing bacteria, and further FMT experiments also confirmed the positive effects of gut microbiome in the vitamin A-treated group (Pang et al., 2021). The supplementation of vitamin A may also directly exert anti-inflammatory functions in the progression of IBD. It has been demonstrated that the intake of vitamin A could increase the levels of NFR-1 and TFAM in both healthy and inflammatory colonic tissues, which supported the potential effects of vitamin A on the preservation of mitochondrial activity (Reifen et al., 2015b). The metabolite of vitamin A, retinoic acid, could also attenuate colonic inflammation and these anti-inflammatory effects have been partially associated with an increase in the expression of IL-22 by $\gamma\delta$ T cells and innate lymphoid cells (Mielke et al., 2013). In addition, the beneficial effects of retinoic acid on IBD have also been connected with the regulation of Treg/Th17 profiles, while the balance and activity of related T cell populations could be regulated by retinoic acid treatment (Bai et al., 2009; Elias et al., 2008).

3.6.2. Vitamin D

Low vitamin D status has been linked with the incidence of various immune systems-related diseases, including IBD (Cantorna and Mahon, 2004). Since there are few natural vitamin D-rich diets, inadequate vitamin D consumption is also problematic. This insufficiency may be more severe in patients with IBD. Vitamin D deficiency has also been regarded as a potential risk involved in the incidence of intestinal inflammation (Levin et al., 2011; Siffledeen et al., 2003; Tajika et al., 2004). In vitamin D receptor-deficient mice, the symptoms of DSS-induced colitis were more severe and further supported the potential crucial roles of vitamin D in the progression of IBD (Kong et al., 2008). Not surprisingly, the supplement of vitamin D in patients with IBD has also been proven as an effective method to improve the disease course. A double-blind clinical study showed that the intake of vitamin D3 (1,200 IU per day) for 12 months could increase the serum levels of vitamin D and decrease the relapse rate of Crohn's disease from 29% to 13% (Jørgensen et al., 2010). Resemble connections between dietary vitamin D and IBD were also revealed by 72,719 women enrolled prospective cohort study, reporting that higher vitamin D status might reduce the risk of IBD (Ananthakrishnan et al., 2012). Numerous studies have been conducted to further examine the direct role of vitamin D in IBD and underlying mechanisms were explored. In IL-10 knockout mice, the deficiency of vitamin D receptor could exacerbate the spontaneous colitis accompanied by a decrease in the number of lymphocytes, highlighting the potential regulation effects of vitamin D in the regulation of immune systems (Froicu et al., 2006). Another study also reported that the treatment of vitamin D3 could improve experimental colitis in mice by regulating the vitamin D receptor-NLRP6 signaling pathway (Gao et al., 2023). Moreover, the positive effects of vitamin D in IBD may also be mediated by gut microbiota. It has been shown that the gut microbiota dysbiosis occurred in vitamin D receptor knockout mice with an increase in *Bacteroidetes* and *Proteobacteria*. The antibiotic treatment further revealed that the aggravated effects of vitamin D deficiency on colitis were mediated by gut microbiota dysbiosis (Ooi et al., 2013).

3.6.3. Vitamin E

The deficiency of vitamin E has also been associated with IBD.

Due to the various symptoms of IBD, including abdominal pain and damage in the intestinal tracts, the intake and absorption of some fat-soluble nutrients can be insufficient and inefficient, especially vitamin E (Fabisiak et al., 2017; Wu et al., 2024). Previous studies indicated that the nutritional risk was significantly higher in adolescents suffering from Crohn's disease with a decrease in vitamin E intake and serum levels (Costa et al., 2015). An increased rate (42.8%) of low serum vitamin E was also shown in severe ulcerative colitis patients, which emphasized the potential connections between vitamin E and IBD (Bousvaros et al., 1998). Some investigations based on humans also supported the positive effects of vitamin E supplementation on IBD. The treatment of d-alpha tocopherol, a vitamin E isomer, could alleviate ulcerative colitis with a decrease in the disease activity index for 64% of patients (Mirbagheri et al., 2008). The anti-inflammatory effects of vitamin E have also been further revealed in animal models. Daily intake of vitamin E (30 U/kg) could suppress the progression of acetic acid-induced colitis in rats with prevented levels of IL-6, MPO, and malondialdehyde (Tahan et al., 2011). Another study also demonstrated that vitamin E could attenuate DSS-induced colitis in mice and further revealed the potential positive effects of vitamin E on DSS-induced barrier damage and gut microbiota dysbiosis (Liu et al., 2021). Although the intake of vitamin E might change the gut microbiota composition of DSS-treated mice with an increase in *Roseburia*, similar effects could not be detected in healthy mice. These studies highlighted the potential roles of gut microbiota in the treatment of vitamin E and some more detailed mechanisms are also required. As a powerful antioxidant, some studies also supported the positive functions of vitamin E in reducing levels of some pro-inflammatory factors in IBD. The administration of tocotrienol could mitigate DSS-induced colitis in mice with a significant decrease in nitric oxide, malondialdehyde, and COX-2, emphasizing the potential anti-oxidative stress effects of vitamin E in IBD (Saw et al., 2019).

3.7. Curcumin

It has been shown that curcumin possesses the extraordinary anti-inflammatory activity (Pituch-Zdanowska et al., 2022). One study based on healthy humans has found that the intake of curcumin might change the gut microbiota composition (Peterson et al., 2018). To further uncover the outcome of curcumin supplements in IBD, some clinical trials have been conducted. In patients with mild-to-moderate ulcerative colitis, the enema treatment of curcumin with the intake of 5-ASA could induce greater improvements compared with patients who received a placebo and 5-ASA (Singla et al., 2014). The comparable effects of curcumin on IBD were also observed during oral treatments. Intake of 3 g of curcumin per day with mesalamine could significantly improve the symptoms of patients with colitis, while 65.3% of patients achieved remission in clinical responses and only 12.5 % in the control group (Lang et al., 2015). Although the underlying mechanisms are still unclear, the intake of curcumin has been regarded as a potential dietary compound to partly relieve IBD without apparent side effects. Based on experimental IBD animal models, the positive effects of curcumin on gut health are further evaluated. Sugimoto et al. (2002) reported that the administration of a diet with curcumin could ameliorate TNBS-induced colitis in mice with the prevention of CD4⁺ T cell infiltration and inflammatory pathways activation. Similar results were also observed in DSS-induced colitis, while the treatment of curcumin could decrease the expression of IL-1 β and suppress NLRP3 inflammasome activation. In this study, the application of the specific NLRP3 inhibitor could abrogate the ef-

fects of curcumin, indicating the potential influence of curcumin on IBD could be associated with the activation of NLRP3 inflammasome (Gong et al., 2018). The interactions between curcumin and gut microbiome have also been declared in animal models. The oral administration of curcumin could improve DSS-induced colitis in mice and change gut microbiota composition with an increase in *Coprococcus*, *Roseburia*, and *Akkermansia* (Guo et al., 2022). However, the exact roles of curcumin-altered gut microbiota are still unknown, and some clinical experiments or FMT are still desired.

3.8. Resveratrol

Resveratrol is a kind of polyphenol dietary component that has attracted much interest with its anti-inflammatory effects and positive influence on the gut microbiome. The potential beneficial impact of resveratrol on IBD has been revealed in previous human clinical trials, the intake of 500 mg of resveratrol for 6 weeks could significantly reduce the plasma levels of TNF- α and activity of NF- κ B in peripheral blood mononuclear cells. The life quality of those patients with active mild-to-moderate ulcerative colitis could be improved, and the clinical disease activity index score was lower in the resveratrol-treated group (Samsami-kor et al., 2015). Moreover, the anti-oxidative effects of resveratrol were also observed in another study, demonstrating that resveratrol treatment in patients could increase the activity of superoxide dismutase and decrease the levels of malondialdehyde in serum (Samsamikor et al., 2016). The positive influence of resveratrol on IBD has also been partially linked with gut microbiome. In DSS-induced colitis mice, resveratrol could improve inflammatory symptoms and repress the expression of some pro-inflammatory cytokines, including GM-CSF, IL-1 β , and IL-6 (Li et al., 2020). Those anti-inflammatory effects have been correlated with the regulation of gut microbiota composition, such as an increase in *Bifidobacterium*. Furthermore, fecal microbiota transplantation has also been performed in another study to reveal the crucial role of resveratrol-altered gut microbiome. Alrafas et al. (2019) found that resveratrol could attenuate TNBS-induced colitis and restore the gut microbiota homeostasis with an increase in *i*-butyric acid. More specifically, this study demonstrated that the resveratrol-altered gut microbiota could promote the amount of CD4⁺FOXP3⁺ T cells and decrease Th17 cells including CD4⁺IFN- γ ⁺ and CD4⁺IL-17⁺ T cells. Another study also highlighted a potential mechanism of resveratrol treatment in colitis, it suggested that resveratrol might regulate the gut microbiome-macrophage-arginine metabolism axis to improve IBD (Xu et al., 2023). The direct anti-inflammatory effects of resveratrol on IBD have also been evaluated in other animal studies, demonstrating that resveratrol could alleviate DSS-induced colitis in mice by regulating the PI3K/Akt/VEGFA pathway (Zhu et al., 2021). Although mechanisms are poorly understood, these animal and human studies have supported that resveratrol could exert beneficial effects on IBD.

4. Gut microbiome-derived lipids in IBD

Lipid metabolites produced by gut microbiota can affect gut homeostasis and influence intestinal inflammation diseases. The lipid content in bacteria accounts for nearly 10% of the dry weight (Brown et al., 2023). Bacteria-derived lipids have long been investigated as structural components, especially in those bacterial membranes. Recent studies revealed that these substances could

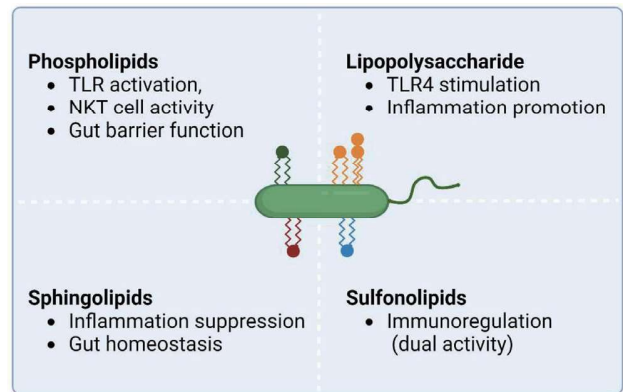


Figure 3. Gut microbiome-derived lipids and their known function in gut homeostasis. Phospholipids are the major components in bacterial membrane, lipopolysaccharide is a component found in Gram-negative bacteria which can activate the immune system through toll-like receptor 4 (TLR4), sphingolipids are produced mostly by commensal *Bacteroidetes* strains, sulfonolipids is present in some gut bacteria like *Bacteroides*, *Alistipes*, and *Flavobacterium* strains. Emerging studies have shown that these gut microbiota-derived lipids play some roles in gut barrier integrity and immunological regulation. Figure 3 was created in Biorender.com (Agreement number: GF27CARL9T).

also be sensed by the immune system cells and regulate various pathways of the host (Chandler and Ernst, 2017). In this section, the possible influence of bacterial lipids on gut health will be discussed (Figure 3).

4.1. Phospholipids

Phospholipids have been known to be the most abundant in bacteria membrane. With the conduction of *Escherichia coli* models, the production and related pathways of phospholipids in bacteria have been uncovered (Zhang and Rock, 2008). It has been suggested that the structure and properties of phospholipids in the membrane could be altered by bacteria to overcome and adapt to a wide range of conditions (Zhang and Rock, 2008). Although the precise mechanisms and connections between gut environment-phospholipids-IBD are still unclear, some studies have already recognized some bacteria-derived phospholipids that could impact the gut health of the host. It has been shown that *Akkermansia muciniphila* could improve experimental colitis in mice by regulating the balance of T cells (Liu et al., 2022). However, some conflict results were also observed in IL-10 knockout mice, showing that *Akkermansia muciniphila* could promote intestinal inflammation (Seregin et al., 2017). Another study recently recognized an immunoregulation-related molecule from *Akkermansia muciniphila*, a diacyl phosphatidylethanolamine with two branched chains (a15:0-i15:0 PE), which could activate TLR2-TLR1 and induce the expression of some inflammatory cytokines, including TNF- α (Bae et al., 2022). Although the data were obtained from *in vitro* models, the potential immunoregulatory effects of bacteria-derived phospholipids on the progression of IBD cannot be ignored. Meanwhile, the regulation role of bacteria-derived phospholipids in gut health and inflammation was also revealed on some pathogens. Compared with mammalian phosphatidylglycerol, *Listeria*-derived phosphatidylglycerol contains shorter and fully-saturated anteiso fatty acid lipid tails, and it can bind to CD1b and be efficiently recognized by natural killer T (NKT) cells to regulate the immunity of the host (Wolf et

al., 2015). NKT cells are a subset of T cells that have also been involved in the progression of IBD based on both clinical trials and murine models (Liao et al., 2013). In addition, previous studies also suggested the potential role of microbiome-derived sphingolipids in the gut barrier possibly by changing the lipid profiles of host cells (Brown et al., 2023).

4.2. Sphingolipids

Some bacteria possess the ability to produce sphingolipids, mainly including *Bacteroidetes*. Deficiency of sphingolipids could induce the alteration of membrane structure, promote susceptibility to oxidative stress, and reduce the survival rate in stressful conditions (An et al., 2011; Heaver et al., 2022). The potential effects of bacteria-derived sphingolipids on the host and intestinal inflammation diseases have also been reported. In patients with IBD, the abundance of *Bacteroidetes* and microbiome-derived sphingolipids were reduced (Brown et al., 2019). In this study, the conduction of germ-free mice and the sphingolipid-deficient bacterial strain showed that the mono-treatment of sphingolipid-deficient bacteria could directly result in intestinal inflammation and gut barrier damage. Combined with the data from *in vitro* studies, microbiome-derived sphingolipids have been recognized as potential components in immune system regulation. Another study has also reported the immune system regulation effects of bacteria-derived sphingolipids. During neonatal development, sphingolipids produced by *Bacteroides fragilis* could impede invariant NKT cell proliferation and then reduce its number in adulthood (An et al., 2014). Moreover, the treatment of bacteria-derived sphingolipids could exert beneficial effects on oxazolone-induced colitis. Although the influence of sphingolipids on immune systems was only observed in the colonic tissues in this study, the potential connections between sphingolipids and the liver functions of the host have also been confirmed. Previous studies also demonstrated that the presence of microbiome-derived sphingolipids could improve fatty liver diseases in animal models, indicating that sphingolipids from bacteria may also have a systemic effect on the host (Le et al., 2022).

4.3. Lipopolysaccharide (LPS)

LPS is a type of bacteria-derived glycolipids that is a crucial component of the membrane bilayers in Gram-negative bacteria. As a barrier element for bacteria, LPS can help to maintain the structure of the membrane and protect them from some detrimental agents, such as antibiotics. In a classic view, LPS is not produced by mammals and thus can activate the innate immune systems, for example, the bacteria-derived LPS can stimulate TLR4 on the host and promote inflammation (Brown et al., 2023). In this regard, LPS has also been regarded as an endotoxin and is associated with numerous inflammatory diseases, including IBD (Candelli et al., 2021). Previous clinical results have shown that the serum levels of LPS and lipopolysaccharide-binding protein were promoted in patients who suffered from IBD compared with healthy individuals, emphasizing the potential pro-inflammatory effects of LPS (Rojo et al., 2006; Tulkens et al., 2020). In mice models, the direct effects of LPS on intestinal inflammation have been examined. It has been suggested that the treatment of LPS could elicit inflammation in the small intestine with body weight loss and increased pro-inflammatory factor expression (Im et al., 2012). Structurally, LPS consists of three domains: lipid A, core oligosaccharide, and O antigen. The immune regulation effects of LPS are mainly dependent

on the lipid A component, and previous studies have found that the structure of lipid A in LPS could be modulated by various bacteria compared to *E. coli* (Okahashi et al., 2021; Park et al., 2009). In this regard, the effects of LPS derived from different bacteria could also be different, and previous *in vitro* studies have supported this speculation. LPS from five species of bacteria could exert different influences on the inflammatory factors' expression and permeability in Caco-2 cells (Stephens and von der Weid, 2020). This species-specific manner of LPS was also observed in mice models. The symptoms of colitis induced by the transfer of CD4⁺CD62L⁺ T cells in *Rag1*^{-/-} mice were more severe with the presence of higher endotoxigenic gut microbiota composition (high levels of *Enterobacteriaceae* and low levels of *Bacteroidetes*) compared with another group with lower endotoxigenic gut microbiota composition (low levels of *Enterobacteriaceae* and high levels of *Bacteroidetes*) (Gronbach et al., 2014). Similarly, it has been suggested that LPS from *Bacteroides* species and *E. coli* could exert different effects on the immune systems of infants (Vatanen et al., 2016), while the LPS from *B. dorei* could inhibit immune activation and inflammatory factors responses to LPS from *E. coli*. These results declared that bacteria-derived LPS could educate and regulate the activation of the immune system, which might also further influence the progression of IBD.

4.4. Sulfonolipids

Sulfonolipids are structurally similar to sphingolipids which is known as a potential immunoregulator factor for the hosts. In the gut microbiota of humans and other animal models, some members of *Bacteroidetes*, including *Alistipes* and *Odoribacter*, can produce sulfonolipids (Walker et al., 2017). Although the mechanisms and connections between bacteria-derived sulfonolipids and IBD are still unclear, some emerging evidence has highlighted the potential immunoregulatory effects of sulfonolipids. In patients with IBD, the abundance of sulfonolipids-produced bacteria was significantly reduced, such as *Alistipes* (Brown et al., 2023; Franzosa et al., 2019). The negative correlation between the abundance of gut microbiota-derived sulfonolipids and IBD has also been directly proven in humans and *Il10*-deficient mice models, which further emphasizes the potential influence of sulfonolipids on IBD (Older et al., 2024). Interestingly, previous *in vitro* studies showed that sulfonolipids could exert dual immunomodulatory activity (Hou et al., 2022; Older et al., 2024). In these studies, the treatment of sulfonolipids could exert mild to moderate pro-inflammatory effects on mouse macrophages, however, sulfonolipids could also suppress the severe inflammatory reaction induced by LPS through TLR signaling. These results clearly show that bacteria-derived sulfonolipid is a potential immunoregulatory factor and may mediate beneficial effects on IBD under specific conditions.

4.5. Short-chain fatty acids (SCFAs)

SCFAs are comprised mostly of acetate (C2), propionate (C3), and butyrate (C4) in an approximate molar ratio of 60:20:20, respectively (Martin-Gallausiaux et al., 2021). The production of SCFAs involves anaerobic fermentation by gut microbiota in the colon. In this process, gut microbiota converts indigestible dietary fibers and other complex polysaccharides into simple sugars, which are further metabolized into SCFAs. Acetate is a net fermentation product for most gut bacteria while propionate and butyrate are produced by more specific bacterial species. Specifically, propionate is limited mainly to the phyla *Bacteroidetes*, some *Firmicutes*

(*Clostridium clusters IX and XI*), and *Actinobacteria* (*Propionibacterium*), and butyrate-producing bacteria belong mainly to the phylum *Firmicutes* (*Clostridium clusters IV and XIVa*) (Louis and Flint, 2017; van der Hee and Wells, 2021). Following their production, SCFAs are absorbed by the colonocytes via specific transporters, and a minor fraction of SCFAs that are not metabolized in the colonocytes are transported to the liver, and circulated throughout the body (Koh et al., 2016).

Previous studies have shown that fecal SCFAs levels are reduced in active IBD (Huda-Faujan et al., 2010). A large body of research also demonstrated that SCFAs play multiple roles in maintaining gut homeostasis. Firstly, butyrate constitutes the major energy source of colonocytes (Ardawi and Newsholme, 1985). In addition, SCFAs play an important role in maintaining gut barrier function by maintaining an environment favorable for commensal bacteria and controlling pathogens' growth. SCFAs, especially butyrate, can regulate the expression of tight junction proteins, which connect adjacent cells in the gut lining, forming a tight seal that prevents the entry of harmful substances. Previous study has shown that butyrate decreases epithelial permeability by regulating the expression of occludin, zonulin and claudins, strengthening the tight junctions and the trans-epithelial resistance *in vitro* (Zheng et al., 2017). Animal-based experiments found that an orally administered butyrate-releasing derivative can improved intestinal epithelial integrity, preventing tight-junction impairment (ZO-1 and occludin) in murine colitis (Simeoli et al., 2017). Moreover, SCFAs, particularly butyrate, modulate the mucus layer thickness and protect the mucosa. In the colon, MUC2 is the predominant mucin glycoprotein produced by the goblet cells. Acetate and butyrate have been shown to stimulate and increase mucin secretion and production in rat colon by up-regulating MUC2 gene expression (Barcelo et al., 2000). Additionally, SCFAs exert anti-inflammatory effects in intestinal mucosa. Propionate and butyrate could suppress NF- κ B reporter activity, immune-related gene expression and cytokine release *in vitro* (Pedersen et al., 2022; Tedelind et al., 2007). Similarly, *in vivo* studies revealed that butyrate and propionate treatment could improve the clinical and inflammatory parameters in murine colitis (Simeoli et al., 2017; Tong et al., 2016). In all, these evidences suggest that supplementation of SCFAs may serve as a promising therapeutic strategy in management of IBD.

5. Future perspective

Dietary and gut microbiome-derived lipids are closely associated with the gut health of humans and can also affect the progression of intestinal inflammatory diseases. Recent studies have arrived at various models and conclusions, and some results were even conflicting and unclear. Therefore, further investigations are still desired to understand the underlying mechanisms for the biological functions of lipids on the GI tracks of host. First, the future effort should further investigate the interactions of various lipid components with gut microbiota in humans. Current studies have studied numerous lipids and examined their influence on inflammatory pathways, mucus layer, and oxidative stress. However, the regulatory mechanism of these lipids on some specific bacterial strains and their clear functions are still understudied. Bacteria-derived lipids, and dietary lipids may also induce the production of bacterial metabolites which may further influence the health of the host. Second, the interactions between lipids and other dietary components should also be subjected to further investigation. Lipid substances have the potential to interact with other dietary components and these interactions may also affect the bioactive

activities of dietary lipids. In conclusion, it is evident that dietary lipids and bacteria-derived lipids can significantly affect the gut microbiota and GI health in the host. The further research is in need to uncover the underlying mechanism by which lipids either from diet or bacteria metabolites modulate the gut health.

Acknowledgments

This work was supported by the Hong Kong Research Grants Council General Research Fund [Project Number CUHK 14104923 and 14102321]

References

- Akerlund, J.E., Björkhem, I., Angelin, B., Liljeqvist, L., and Einarsson, K. (1994). Apparent selective bile acid malabsorption as a consequence of ileal exclusion: effects on bile acid, cholesterol, and lipoprotein metabolism. *Gut* 35(8): 1116–1120.
- Alexander, M., Ang, Q.Y., Nayak, R.R., Bustion, A.E., Sandy, M., Zhang, B., Upadhyay, V., Pollard, K.S., Lynch, S.V., and Turnbaugh, P.J. (2022). Human gut bacterial metabolism drives Th17 activation and colitis. *Cell Host Microbe* 30(1): 17–30.e19.
- Alrafay, H.R., Busbee, P.B., Nagarkatti, M., and Nagarkatti, P.S. (2019). Resveratrol modulates the gut microbiota to prevent murine colitis development through induction of Tregs and suppression of Th17 cells. *J. Leukoc. Biol.* 106(2): 467–480.
- Alsharairi, N.A. (2022). The Therapeutic Role of Short-Chain Fatty Acids Mediated Very Low-Calorie Ketogenic Diet–Gut Microbiota Relationships in Paediatric Inflammatory Bowel Diseases. *Nutrients* 14(19): 4113.
- An, D., Na, C., Bielawski, J., Hannun, Y.A., and Kasper, D.L. (2011). Membrane sphingolipids as essential molecular signals for *Bacteroides* survival in the intestine. *Proc. Natl. Acad. Sci. U.S.A.* 108(Suppl 1): 4666–4671.
- An, D., Oh, S.F., Olszak, T., Neves, J.F., Avci, F.Y., Erturk-Hasdemir, D., Lu, X., Zeissig, S., Blumberg, R.S., and Kasper, D.L. (2014). Sphingolipids from a Symbiotic Microbe Regulate Homeostasis of Host Intestinal Natural Killer T Cells. *Cell* 156(1): 123–133.
- Ananthakrishnan, A.N. (2015). Epidemiology and risk factors for IBD. *Nat. Rev. Gastroenterol. Hepatol.* 12(4): 205–217.
- Ananthakrishnan, A.N., Khalili, H., Higuchi, L.M., Bao, Y., Korzenik, J.R., Giovannucci, E.L., Richter, J.M., Fuchs, C.S., and Chan, A.T. (2012). Higher Predicted Vitamin D Status Is Associated With Reduced Risk of Crohn's Disease. *Gastroenterology* 142(3): 482–489.
- Ananthakrishnan, A.N., Khalili, H., Konijeti, G.G., Higuchi, L.M., de Silva, P., Fuchs, C.S., Willett, W.C., Richter, J.M., and Chan, A.T. (2014). Long-term intake of dietary fat and risk of ulcerative colitis and Crohn's disease. *Gut* 63(5): 776–784.
- Anderson, R.C., Cookson, A.L., McNabb, W.C., Park, Z., McCann, M.J., Kelly, W.J., and Roy, N.C. (2010). *Lactobacillus plantarum* MB452 enhances the function of the intestinal barrier by increasing the expression levels of genes involved in tight junction formation. *BMC Microbiol.* 10: 1–11.
- Ang, Q.Y., Alexander, M., Newman, J.C., Tian, Y., Cai, J., Upadhyay, V., Turnbaugh, J.A., Verdin, E., Hall, K.D., Leibel, R.L., Ravussin, E., Rosenbaum, M., Patterson, A.D., and Turnbaugh, P.J. (2020). Ketogenic Diets Alter the Gut Microbiome Resulting in Decreased Intestinal Th17 Cells. *Cell* 181(6): 1263–1275.e1216.
- Anunziato, F., Cosmi, L., Santarlasci, V., Maggi, L., Liotta, F., Mazzinghi, B., Parente, E., Fili, L., Ferri, S., Frosali, F., Giudici, F., Romagnani, P., Parronchi, P., Tonelli, F., Maggi, E., and Romagnani, S. (2007). Phenotypic and functional features of human Th17 cells. *J. Exp. Med.* 204(8): 1849–1861.
- Ardawi, M.S., and Newsholme, E.A. (1985). Fuel utilization in colonocytes of the rat. *Biochem. J.* 231(3): 713–719.
- Armand, M. (2007). Lipases and lipolysis in the human digestive tract:

- where do we stand? *Curr. Opin. Clin. Nutr. Metab. Care* 10(2): 156–164.
- Astore, C., Nagpal, S., and Gibson, G. (2022). Mendelian Randomization Indicates a Causal Role for Omega-3 Fatty Acids in Inflammatory Bowel Disease. *Int. J. Mol. Sci.* 23(22): 14380.
- Bae, M., Cassilly, C.D., Liu, X., Park, S.M., Tusi, B.K., Chen, X., Kwon, J., Filipčík, P., Bolze, A.S., Liu, Z., Vlamakis, H., Graham, D.B., Buhrlage, S.J., Xavier, R.J., and Clardy, J. (2022). Akkermansia muciniphila phospholipid induces homeostatic immune responses. *Nature* 608(7921): 168–173.
- Bai, A., Lu, N., Guo, Y., Liu, Z., Chen, J., and Peng, Z. (2009). All-trans retinoic acid down-regulates inflammatory responses by shifting the Treg/Th17 profile in human ulcerative and murine colitis. *J. Leukocyte Biol.* 86(4): 959–969.
- Bai, B., Yamamoto, K., Sato, H., Sugiura, H., and Tanaka, T. (2005). Combined Effect of 25-Hydroxycholesterol and IL-1 β on IL-8 Production in Human Colon Carcinoma Cell Line (Caco-2). *Inflammation* 29(4): 141–146.
- Barcelo, A., Claustre, J., Moro, F., Chayvialle, J.A., Cuber, J.C., and Plaisancie, P. (2000). Mucin secretion is modulated by luminal factors in the isolated vascularly perfused rat colon. *Gut* 46(2): 218–224.
- Barrasa, J.I., Olmo, N., Pérez-Ramos, P., Santiago-Gómez, A., Lecona, E., Turnay, J., and Antonia Lizarbe, M. (2011). Deoxycholic and chenodeoxycholic bile acids induce apoptosis via oxidative stress in human colon adenocarcinoma cells. *Apoptosis* 16: 1054–1067.
- Bassaganya-Riera, J., Hontecillas, R., Horne, W.T., Sandridge, M., Herfarth, H.H., Bloomfield, R., and Isaacs, K.L. (2012). Conjugated linoleic acid modulates immune responses in patients with mild to moderately active Crohn's disease. *Clin. Nutr.* 31(5): 721–727.
- Bassaganya-Riera, J., Reynolds, K., Martino-Catt, S., Cui, Y., Hennighausen, L., Gonzalez, F., Rohrer, J., Benninghoff, A.U., and Hontecillas, R. (2004). Activation of PPAR γ and δ by conjugated linoleic acid mediates protection from experimental inflammatory bowel disease. *Gastroenterology* 127(3): 777–791.
- Bennike, T.B., Carlsen, T.G., Ellingsen, T., Bonderup, O.K., Glerup, H., Bøgsted, M., Christiansen, G., Birkelund, S., Stensballe, A., and Andersen, V. (2015). Neutrophil Extracellular Traps in Ulcerative Colitis: A Proteome Analysis of Intestinal Biopsies. *Inflammatory Bowel Dis.* 21(9): 2052–2067.
- Bergstrom, K., Shan, X., Casero, D., Batushansky, A., Lagishetty, V., Jacobs, J.P., Hoover, C., Kondo, Y., Shao, B., Gao, L., Zandberg, W., Noyovitz, B., McDaniel, J.M., Gibson, D.L., Pakpour, S., Kazemian, N., McGee, S., Houchen, C.W., Rao, C.V., Griffin, T.M., Sonnenburg, J.L., McEver, R.P., Braun, J., and Xia, L. (2020). Proximal colon-derived O-glycosylated mucus encapsulates and modulates the microbiota. *Science* 370(6515): 467–472.
- Bian, X., Wu, W., Yang, L., Lv, L., Wang, Q., Li, Y., Ye, J., Fang, D., Wu, J., Jiang, X., Shi, D., and Li, L. (2019). Administration of Akkermansia muciniphila Ameliorates Dextran Sulfate Sodium-Induced Ulcerative Colitis in Mice. *Front. Microbiol.* 10: 2259.
- Biasi, F., Mascia, C., Astegiano, M., Chiarpotto, E., Nano, M., Vizio, B., Leonarduzzi, G., and Poli, G. (2009). Pro-oxidant and proapoptotic effects of cholesterol oxidation products on human colonic epithelial cells: a potential mechanism of inflammatory bowel disease progression. *Free Radic. Biol. Med.* 47: 1731–1741.
- Bousvaros, A., Zurakowski, D., Duggan, C., Law, T., Rifai, N., Goldberg, N.E., and Leichtner, A.M. (1998). Vitamins A and E Serum Levels in Children and Young Adults with Inflammatory Bowel Disease: Effect of Disease Activity. *J. Pediatr. Gastroenterol. Nutr.* 26(2): 129–135.
- Brown, E.M., Clardy, J., and Xavier, R.J. (2023). Gut microbiome lipid metabolism and its impact on host physiology. *Cell Host Microbe* 31(2): 173–186.
- Brown, E.M., Ke, X., Hitchcock, D., Jeanfavre, S., Avila-Pacheco, J., Nakata, T., Arthur, T.D., Fornelos, N., Heim, C., Franzosa, E.A., Watson, N., Huttenhower, C., Haiser, H.J., Dillow, G., Graham, D.B., Finlay, B.B., Kostic, A.D., Porter, J.A., Vlamakis, H., Clish, C.B., and Xavier, R.J. (2019). Bacteroides-Derived Sphingolipids Are Critical for Maintaining Intestinal Homeostasis and Symbiosis. *Cell Host Microbe* 25: 668–680.e667.
- Bruce, A., Black, M., and Bhattacharya, S. (2014). Mode of Delivery and Risk of Inflammatory Bowel Disease in the Offspring: Systematic Review and Meta-analysis of Observational Studies. *Inflammatory Bowel Dis.* 20(7): 1217–1226.
- Caetano, M.A.F., and Castelucci, P. (2022). Role of short chain fatty acids in gut health and possible therapeutic approaches in inflammatory bowel diseases. *World J. Clin. Cases* 10(28): 9985–10003.
- Cai, J., Sun, L., and Gonzalez, F.J. (2022). Gut microbiota-derived bile acids in intestinal immunity, inflammation, and tumorigenesis. *Cell Host Microbe* 30(3): 289–300.
- Camoglio, L., Te Velde, A.A., Tigges, A.J., Das, P.K., and Van Deventer, S.J.H. (1998). Altered Expression of Interferon- γ and Interleukin-4 in Inflammatory Bowel Disease. *Inflammatory Bowel Dis.* 4(4): 285–290.
- Candelli, M., Franza, L., Pignataro, G., Ojetti, V., Covino, M., Piccioni, A., Gasbarrini, A., and Franceschi, F. (2021). Interaction between Lipopolysaccharide and Gut Microbiota in Inflammatory Bowel Diseases. *Int. J. Mol. Sci.* 22(12): 6242.
- Cani, P.D., Bibiloni, R., Knauf, C., Waget, A., Neyrinck, A.M., Delzenne, N.M., and Burcelin, R. (2008). Changes in gut microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity and diabetes in mice. *Diabetes* 57(6): 1470–1481.
- Cani, P.D., Neyrinck, A.M., Fava, F., Knauf, C., Burcelin, R.G., Tuohy, K.M., Gibson, G.R., and Delzenne, N.M. (2007). Selective increases of bifidobacteria in gut microflora improve high-fat-diet-induced diabetes in mice through a mechanism associated with endotoxaemia. *Diabetologia* 50: 2374–2383.
- Cantorna, M.T., and Mahon, B.D. (2004). Mounting Evidence for Vitamin D as an Environmental Factor Affecting Autoimmune Disease Prevalence. *Exp. Biol. Med.* 229(11): 1136–1142.
- Caprioli, F., Pallone, F., and Monteleone, G. (2008). Th17 immune response in IBD: A new pathogenic mechanism. *J. Crohn's Colitis* 2(4): 291–295.
- Cha, H.R., Chang, S.Y., Chang, J.H., Kim, J.O., Yang, J.Y., Kim, C.H., and Kweon, M.N. (2010). Downregulation of Th17 cells in the small intestine by disruption of gut flora in the absence of retinoic acid. *J. Immunol.* 184(12): 6799–6806.
- Chalubinski, M., Wojdan, K., Gorzelak, P., Borowiec, M., and Broncel, M. (2014). The effect of oxidized cholesterol on barrier functions and IL-10 mRNA expression in human intestinal epithelium co-cultured with dendritic cells in the transwell system. *Food Chem. Toxicol.* 69: 289–293.
- Chandler, C.E., and Ernst, R.K. (2017). Bacterial lipids: powerful modifiers of the innate immune response. *F1000Res.* 6: F1000.
- Chassaing, B., and Darfeuille-Michaud, A. (2011). The Commensal Microbiota and Enteropathogens in the Pathogenesis of Inflammatory Bowel Diseases. *Gastroenterology* 140(6): 1720–1728.e1723.
- Chelakkot, C., Choi, Y., Kim, D.K., Park, H.T., Ghim, J., Kwon, Y., Jeon, J., Kim, M.S., Jee, Y.K., Gho, Y.S., Park, H.S., Kim, Y.K., and Ryu, S.H. (2018a). Akkermansia muciniphila-derived extracellular vesicles influence gut permeability through the regulation of tight junctions. *Exp. Mol. Med.* 50(2): e450–e450.
- Chelakkot, C., Ghim, J., and Ryu, S.H. (2018b). Mechanisms regulating intestinal barrier integrity and its pathological implications. *Exp. Mol. Med.* 50(8): 1–9.
- Chen, Y., Gao, H., Zhao, J., Ross, R.P., Stanton, C., Zhang, H., Chen, W., and Yang, B. (2023a). Exploiting lactic acid bacteria for inflammatory bowel disease: A recent update. *Trends Food Sci. Technol.* 138: 126–140.
- Chen, Y., Mai, Q., Chen, Z., Lin, T., Cai, Y., Han, J., Wang, Y., Zhang, M., Tan, S., Wu, Z., Chen, L., Zhang, Z., Yang, Y., Cui, T., Ouyang, B., Sun, Y., Yang, L., Xu, L., Zhang, S., Li, J., Shen, H., Liu, L., Zeng, L., Zhang, S., and Zeng, G. (2023b). Dietary palmitoleic acid reprograms gut microbiota and improves biological therapy against colitis. *Gut Microbes* 15: 2211501.
- Chen, Y., Yang, B., Ross, R.P., Jin, Y., Stanton, C., Zhao, J., Zhang, H., and Chen, W. (2019). Orally Administered CLA Ameliorates DSS-Induced Colitis in Mice via Intestinal Barrier Improvement, Oxidative Stress Reduction, and Inflammatory Cytokine and Gut Microbiota Modulation. *J. Agric. Food Chem.* 67(48): 13282–13298.
- Çimen, I., Kocatürk, B., Koyuncu, S., Tufanlı, Ö., Onat, U.I., Yıldırım, A.D., Apaydın, O., Demirsoy, Ş., Aykut, Z.G., Nguyen, U.T., Watkins, S.M., Hotamışgılı, G.S., and Erbay, E. (2016). Prevention of atherosclerosis by bioactive palmitoleate through suppression of organelle stress and inflammasome activation. *Sci. Transl. Med.* 8(358): 358ra126.

- Collins, S.L., Stine, J.G., Bisanz, J.E., Okafor, C.D., and Patterson, A.D. (2023). Bile acids and the gut microbiota: Metabolic interactions and impacts on disease. *Nat. Rev. Microbiol.* 21(4): 236–247.
- Costa, C.O.P.C., Carrilho, F.J., Nunes, V.S., Sipahi, A.M., and Rodrigues, M. (2015). A snapshot of the nutritional status of Crohn's disease among adolescents in Brazil: a prospective cross-sectional study. *BMC Gastroenterol.* 15(1): 172.
- de La Serre, C.B., Ellis, C.L., Lee, J., Hartman, A.L., Rutledge, J.C., and Raybould, H.E. (2010). Propensity to high-fat diet-induced obesity in rats is associated with changes in the gut microbiota and gut inflammation. *Am. J. Physiol. Gastrointest. Liver Physiol.* 299(2): G440–G448.
- De Preter, V., Machiels, K., Joossens, M., Arijis, I., Matthys, C., Vermeire, S., Rutgeerts, P., and Verbeke, K. (2015). Faecal metabolite profiling identifies medium-chain fatty acids as discriminating compounds in IBD. *Gut* 64(3): 447–458.
- de Silva, P.S., Luben, R., Shrestha, S.S., Khaw, K.T., and Hart, A.R. (2014). Dietary arachidonic and oleic acid intake in ulcerative colitis etiology: a prospective cohort study using 7-day food diaries. *Eur. J. Gastroenterol. Hepatol.* 26(1): 11–18.
- de Souza, H.S.P., and Focchi, C. (2016). Immunopathogenesis of IBD: current state of the art. *Nat. Rev. Gastroenterol. Hepatol.* 13(1): 13–27.
- Deiana, M., Calfapietra, S., Incani, A., Atzeri, A., Rossin, D., Loi, R., Sottero, B., Iaia, N., Poli, G., and Biasi, F. (2017). Derangement of intestinal epithelial cell monolayer by dietary cholesterol oxidation products. *Free Radic. Biol. Med.* 113: 539–550.
- Deng, C., Li, M., Liu, Y., Yan, C., He, Z., Chen, Z.Y., and Zhu, H. (2023). Cholesterol Oxidation Products: Potential Adverse Effect and Prevention of Their Production in Foods. *J. Agric. Food Chem.* 71(48): 18645–18659.
- Deol, P., Ruegger, P., Logan, G.D., Shawki, A., Li, J., Mitchell, J.D., Yu, J., Piamthai, V., Radi, S.H., Hasnain, S., Borkowski, K., Newman, J.W., McCole, D.F., Nair, M.G., Hsiao, A., Borneman, J., and Sladek, F.M. (2023). Diet high in linoleic acid dysregulates the intestinal endocannabinoid system and increases susceptibility to colitis in Mice. *Gut Microbes* 15(1): 2229945.
- Devkota, S., Wang, Y., Musch, M.W., Leone, V., Fehlner-Peach, H., Nadimpalli, A., Antonopoulos, D.A., Jabri, B., and Chang, E.B. (2012). Dietary-fat-induced taurocholic acid promotes pathobiont expansion and colitis in *Il10^{-/-}* mice. *Nature* 487(7405): 104–108.
- Dheer, R., Santaolalla, R., Davies, J.M., Lang, J.K., Phillips, M.C., Pastorini, C., Vazquez-Pertejo, M.T., and Abreu, M.T. (2016). Intestinal Epithelial Toll-Like Receptor 4 Signaling Affects Epithelial Function and Colonic Microbiota and Promotes a Risk for Transmissible Colitis. *Infect. Immun.* 84(3): 798–810.
- Dinallo, V., Marafini, I., Di Fusco, D., Laudisi, F., Franzè, E., Di Grazia, A., Figliuzzi, M.M., Caprioli, F., Stolfi, C., Monteleone, I., and Monteleone, G. (2019). Neutrophil Extracellular Traps Sustain Inflammatory Signals in Ulcerative Colitis. *J. Crohns. Colitis.* 13(6): 772–784.
- Ding, S., Chi, M.M., Scull, B.P., Rigby, R., Schwerbrock, N.M., Magnus, S., Jobin, C., and Lund, P.K. (2010). High-Fat Diet: Bacteria Interactions Promote Intestinal Inflammation Which Precedes and Correlates with Obesity and Insulin Resistance in Mouse. *PLoS One* 5(8): e12191.
- Dominguez-Bello, M.G., Blaser, M.J., Ley, R.E., and Knight, R. (2011). Development of the human gastrointestinal microbiota and insights from high-throughput sequencing. *Gastroenterology* 140(6): 1713–1719.
- Dong, Y., Huang, C., Yang, J., Zheng, Z., and Dai, Z. (2022). Docosapentaenoic Acid (DPA, 22:5 n-3) Alleviates Ulcerative Colitis via Modification of Gut Microbiota and Their Metabolism. *Nutrients* 14(19): 4204.
- Du, Q., Wang, Q., Fan, H., Wang, J., Liu, X., Wang, H., Wang, Y., and Hu, R. (2016). Dietary cholesterol promotes AOM-induced colorectal cancer through activating the NLRP3 inflammasome. *Biochem. Pharmacol.* 105: 42–54.
- Edelblum, K.L., and Turner, J.R. (2009). The tight junction in inflammatory disease: communication breakdown. *Curr. Opin. Pharmacol.* 9(6): 715–720.
- Eichmann, F., Sellem, L., Wittenbecher, C., Jäger, S., Kuxhaus, O., Prada, M., Cuadrat, R., Jackson, K.G., Lovegrove, J.A., and Schulze, M.B. (2022). Deep Lipidomics in Human Plasma: Cardiometabolic Disease Risk and Effect of Dietary Fat Modulation. *Circulation* 146(1): 21–35.
- Elias, K.M., Laurence, A., Davidson, T.S., Stephens, G., Kanno, Y., Shevach, E.M., and O'Shea, J.J. (2008). Retinoic acid inhibits Th17 polarization and enhances FoxP3 expression through a Stat-3/Stat-5 independent signaling pathway. *Blood* 111(3): 1013–1020.
- Fabisiak, N., Fabisiak, A., Watala, C., and Fichna, J. (2017). Fat-soluble Vitamin Deficiencies and Inflammatory Bowel Disease: Systematic Review and Meta-Analysis. *J. Clin. Gastroenterol.* 51(10): 878–889.
- Fernández, J., de la Fuente, V.G., García, M.T.F., Sánchez, J.G., Redondo, B.I., Villar, C.J., and Lombó, F. (2020). A diet based on cured acorn-fed ham with oleic acid content promotes anti-inflammatory gut microbiota and prevents ulcerative colitis in an animal model. *Lipids Health Dis.* 19(1): 28.
- Ferrand, A., Al Nabhani, Z., Tapias, N.S., Mas, E., Hugot, J.P., and Barreau, F. (2019). NOD2 Expression in Intestinal Epithelial Cells Protects Toward the Development of Inflammation and Associated Carcinogenesis. *Cell. Mol. Gastroenterol. Hepatol.* 7(2): 357–369.
- Franzosa, E.A., Sirota-Madi, A., Avila-Pacheco, J., Fornelos, N., Haiser, H.J., Reinker, S., Vatanen, T., Hall, A.B., Mallick, H., McIver, L.J., Sauk, J.S., Wilson, R.G., Stevens, B.W., Scott, J.M., Pierce, K., Deik, A.A., Bullock, K., Imhann, F., Porter, J.A., Zhernakova, A., Fu, J., Weersma, R.K., Wijmenga, C., Clish, C.B., Vlamakis, H., Huttenhower, C., and Xavier, R.J. (2019). Gut microbiome structure and metabolic activity in inflammatory bowel disease. *Nat. Microbiol.* 4(2): 293–305.
- Froicu, M., Zhu, Y., and Cantorna, M.T. (2006). Vitamin D receptor is required to control gastrointestinal immunity in IL-10 knockout mice. *Immunology* 117(3): 310–318.
- Furusawa, Y., Obata, Y., Fukuda, S., Endo, T.A., Nakato, G., Takahashi, D., Nakanishi, Y., Uetake, C., Kato, K., Kato, T., Takahashi, M., Fukuda, N.N., Murakami, S., Miyachi, E., Hino, S., Atarashi, K., Onawa, S., Fujimura, Y., Lockett, T., Clarke, J.M., Topping, D.L., Tomita, M., Hori, S., Ohara, O., Morita, T., Koseki, H., Kikuchi, J., Honda, K., Hase, K., and Ohno, H. (2013). Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* 504(7480): 446–450.
- Gao, H., Zhou, H., Zhang, Z., Gao, J., Li, J., and Li, X. (2023). Vitamin D3 alleviates inflammation in ulcerative colitis by activating the VDR-NLRP6 signaling pathway. *Front. Immunol.* 14: 1135930.
- Gao, Q., Esworthy, S.R., Kim, B.-W., Synold, T.W., Smith, D.D., and Chu, F.-F. (2010). Atherogenic diets exacerbate colitis in mice deficient in glutathione peroxidase. *Inflammatory Bowel Dis.* 16(12): 2043–2054.
- Gassull, M.A., Fernández-Bañares, F., Cabré, E., Papo, M., Gaffner, M.H., Sánchez-Lombrana, J.L., Richart, C., Malchow, H., González-Huix, F., Esteve, M., and European Group on Enteral Nutrition in Crohn's Disease. (2002). Fat composition may be a clue to explain the primary therapeutic effect of enteral nutrition in Crohn's disease: results of a double blind randomised multicentre European trial. *Gut* 51(2): 164–168.
- GBD 2017 Inflammatory Bowel Disease Collaborators. (2017). The global, regional, and national burden of inflammatory bowel disease in 195 countries and territories, 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet Gastroenterol. Hepatol.* 5(1): 17–30.
- Gentile, C.L., and Weir, T.L. (2018). The gut microbiota at the intersection of diet and human health. *Science* 362(6416): 776–780.
- Ghezzal, S., Postal, B.G., Quevrain, E., Brot, L., Seksik, P., Leturque, A., Thenet, S., and Carriere, V. (2020). Palmitic acid damages gut epithelium integrity and initiates inflammatory cytokine production. *Biochim. Biophys. Acta Mol. Cell. Biol. Lipids* 1865: 158530.
- Gitter, A.H., Wullstein, F., Fromm, M., and Schulzke, J.D. (2001). Epithelial barrier defects in ulcerative colitis: Characterization and quantification by electrophysiological imaging. *Gastroenterology* 121(6): 1320–1328.
- Gomez-Bris, R., Saez, A., Herrero-Fernandez, B., Rius, C., Sanchez-Martinez, H., and Gonzalez-Granado, J.M. (2023). CD4 T-Cell Subsets and the Pathophysiology of Inflammatory Bowel Disease. *Int. J. Mol. Sci.* 24(3): 2696.
- Gong, Z., Zhao, S., Zhou, J., Yan, J., Wang, L., Du, X., Li, H., Chen, Y., Cai, W., and Wu, J. (2018). Curcumin alleviates DSS-induced colitis via inhibiting NLRP3 inflammasome activation and IL-1 β production. *Mol. Immunol.* 104: 11–19.
- Gronbach, K., Flade, I., Holst, O., Lindner, B., Ruscheweyh, H.J., Wittmann, A., Menz, S., Schwierz, A., Adam, P., Stecher, B., Josenhans, C., Suer-

- baum, S., Gruber, A.D., Kulik, A., Huson, D., Autenrieth, I.B., and Frick, J.S. (2014). Endotoxicity of Lipopolysaccharide as a Determinant of T-Cell-Mediated Colitis Induction in Mice. *Gastroenterology* 146(3): 765–775.
- Guina, T., Deiana, M., Calfapietra, S., Cabboi, B., Maina, M., Tuberoso, C.I., Leonarduzzi, G., Gamba, P., Gargiulo, S., Testa, G., Poli, G., and Biasi, F. (2015). The role of p38 MAPK in the induction of intestinal inflammation by dietary oxysterols: modulation by wine phenolics. *Food Funct.* 6(4): 1218–1228.
- Gulhane, M., Murray, L., Lourie, R., Tong, H., Sheng, Y.H., Wang, R., Kang, A., Schreiber, V., Wong, K.Y., Magor, G., Denman, S., Begun, J., Florin, T.H., Perkins, A., Cuiv, P.O., McGuckin, M.A., and Hasnain, S.Z. (2016). High Fat Diets Induce Colonic Epithelial Cell Stress and Inflammation that is Reversed by IL-22. *Sci. Rep.* 6(1): 28990.
- Gunasekera, D.C., Ma, J., Vacharathit, V., Shah, P., Ramakrishnan, A., Uprety, P., Shen, Z., Sheh, A., Brayton, C.F., Whary, M.T., Fox, J.G., and Bream, J.H. (2020). The development of colitis in *Il10^{-/-}* mice is dependent on IL-22. *Mucosal Immunol.* 13(3): 493–506.
- Guo, S., Al-Sadi, R., Said, H.M., and Ma, T.Y. (2013). Lipopolysaccharide causes an increase in intestinal tight junction permeability in vitro and in vivo by inducing enterocyte membrane expression and localization of TLR-4 and CD14. *Am. J. Pathol.* 182(2): 375–387.
- Guo, X., Xu, Y., Geng, R., Qiu, J., and He, X. (2022). Curcumin Alleviates Dextran Sulfate Sodium-Induced Colitis in Mice Through Regulating Gut Microbiota. *Mol. Nutr. Food Res.* 66(8): e2100943.
- Hall, K.D., and Chung, S.T. (2018). Low-carbohydrate diets for the treatment of obesity and type 2 diabetes. *Curr. Opin. Clin. Nutr. Metab. Care* 21(4): 308–312.
- Hao, W., Kwek, E., He, Z., Zhu, H., Liu, J., Zhao, Y., Ma, K.Y., He, W.S., and Chen, Z.Y. (2020). Ursolic acid alleviates hypercholesterolemia and modulates the gut microbiota in hamsters. *Food Funct.* 11(7): 6091–6103.
- Hao, W., He, Z., Zhu, H., Liu, J., Kwek, E., Zhao, Y., Ma, K.Y., He, W.S., and Chen, Z.Y. (2019). Sea buckthorn seed oil reduces blood cholesterol and modulates gut microbiota. *Food Funct.* 10(9): 5669–5681.
- Hart, A.L., Al-Hassi, H.O., Rigby, R.J., Bell, S.J., Emmanuel, A.V., Knight, S.C., Kamm, M.A., and Stagg, A.J. (2005). Characteristics of intestinal dendritic cells in inflammatory bowel diseases. *Gastroenterology* 129(1): 50–65.
- Hartwig, O., Shetab Boushehri, M.A., Shalaby, K.S., Loretz, B., Lamprecht, A., and Lehr, C.-M. (2021). Drug delivery to the inflamed intestinal mucosa – targeting technologies and human cell culture models for better therapies of IBD. *Adv. Drug Delivery Rev.* 175: 113828.
- Haskey, N., Ye, J., Estaki, M., Verdugo Meza, A.A., Barnett, J.A., Yousefi, M., Birnie, B.W., Gruenheid, S., Ghosh, S., and Gibson, D.L. (2022). A Mediterranean-like fat blend protects against the development of severe colitis in the mucin-2 deficient murine model. *Gut Microbes* 14(1): 2055441.
- Heaver, S.L., Le, H.H., Tang, P., Baslé, A., Mirretta Barone, C., Vu, D.L., Waters, J.L., Marles-Wright, J., Johnson, E.L., Campopiano, D.J., and Ley, R.E. (2022). Characterization of inositol lipid metabolism in gut-associated Bacteroidetes. *Nat. Microbiol.* 7(7): 986–1000.
- Hildebrandt, M.A., Hoffmann, C., Sherrill-Mix, S.A., Keilbaugh, S.A., Hamady, M., Chen, Y.Y., Knight, R., Ahima, R.S., Bushman, F., and Wu, G.D. (2009). High-fat diet determines the composition of the murine gut microbiome independently of obesity. *Gastroenterology* 137(5): 1716–1724.e1712.
- Hooper, L.V., Stappenbeck, T.S., Hong, C.V., and Gordon, J.I. (2003). Angiogenins: a new class of microbicidal proteins involved in innate immunity. *Nat. Immunol.* 4(3): 269–273.
- Hosomi, K., Kiyono, H., and Kunisawa, J. (2020). Fatty acid metabolism in the host and commensal bacteria for the control of intestinal immune responses and diseases. *Gut Microbes* 11(3): 276–284.
- Hou, L., Tian, H.Y., Wang, L., Ferris, Z.E., Wang, J., Cai, M., Older, E.A., Raja, M.R.K., Xue, D., Sun, W., Nagarkatti, P., Nagarkatti, M., Chen, H., Fan, D., Tang, X., and Li, J. (2022). Identification and Biosynthesis of Pro-Inflammatory Sulfonolipids from an Opportunistic Pathogen *Chryseobacterium gleum*. *ACS Chem. Biol.* 17(5): 1197–1206.
- Hsieh, C.Y., Osaka, T., Moriyama, E., Date, Y., Kikuchi, J., and Tsuneda, S. (2015). Strengthening of the intestinal epithelial tight junction by *Bifidobacterium bifidum*. *Physiol. Rep.* 3(3): e12327.
- Hu, J., La Vecchia, C., de Groh, M., Negri, E., Morrison, H., and Mery, L. (2012). Dietary cholesterol intake and cancer. *Ann. Oncol.* 23(2): 491–500.
- Huda-Faujan, N., Abdulmir, A.S., Fatimah, A.B., Anas, O.M., Shuhaimi, M., Yazid, A.M., and Loong, Y.Y. (2010). The impact of the level of the intestinal short chain Fatty acids in inflammatory bowel disease patients versus healthy subjects. *Open Biochem. J.* 4: 53–58.
- Hudert, C.A., Weylandt, K.H., Lu, Y., Wang, J., Hong, S., Dignass, A., Serhan, C.N., and Kang, J.X. (2006). Transgenic mice rich in endogenous omega-3 fatty acids are protected from colitis. *Proc. Natl. Acad. Sci.* 103(30): 11276–11281.
- IBD in EPIC Study Investigators, Tjonneland, A., Overvad, K., Bergmann, M.M., Nagel, G., Linseisen, J., Hallmans, G., Palmqvist, R., Sjodin, H., Hagglund, G., Berglund, G., Lindgren, S., Grip, O., Palli, D., Day, N.E., Khaw, K.T., Bingham, S., Riboli, E., Kennedy, H., and Hart, A. (2009). Linoleic acid, a dietary n-6 polyunsaturated fatty acid, and the aetiology of ulcerative colitis: a nested case-control study within a European prospective cohort study. *Gut* 58(12): 1606–1611.
- Im, E., Riegler, F.M., Pothoulakis, C., and Olson, M.R. (2012). Elevated lipopolysaccharide in the colon evokes intestinal inflammation, aggravated in immune modulator-impaired mice. *Am. J. Physiol. Gastrointest. Liver Physiol.* 303(4): G490–G497.
- Imam, T., Park, S., Kaplan, M.H., and Olson, M.R. (2018). Effector T Helper Cell Subsets in Inflammatory Bowel Diseases. *Front. Immunol.* 9: 1212.
- Jia, M., Zhang, Y., Gao, Y., and Ma, X. (2020a). Effects of medium chain fatty acids on intestinal health of monogastric animals. *Curr. Protein Pept. Sci.* 21(8): 777–784.
- Jia, W., Rajani, C., Xu, H., and Zheng, X. (2020b). Gut microbiota alterations are distinct for primary colorectal cancer and hepatocellular carcinoma. *Protein Cell* 12(5): 374–393.
- Jiang, J., Chen, L., Sun, R., Yu, T., Jiang, S., and Chen, H. (2023). Characterization of serum polyunsaturated fatty acid profile in patients with inflammatory bowel disease. *Ther. Adv. Chronic Dis.* 14: 20406223231156826.
- Johansson, M.E.V., and Hansson, G.C. (2016). Immunological aspects of intestinal mucus and mucins. *Nat. Rev. Immunol.* 16(10): 639–649.
- Johansson, M.E., Sjövall, H., and Hansson, G.C. (2013). The gastrointestinal mucus system in health and disease. *Nat. Rev. Gastroenterol. Hepatol.* 10(6): 352–361.
- Jørgensen, S.P., Agnholt, J., Glerup, H., Lyhne, S., Villadsen, G.E., Hvas, C.L., Bartels, L.E., Kelsen, J., Christensen, L.A., and Dahlerup, J.F. (2010). Clinical trial: vitamin D3 treatment in Crohn's disease – a randomized double-blind placebo-controlled study. *Aliment. Pharmacol. Ther.* 32(3): 377–383.
- Kaur, H., Kaur, G., and Ali, S.A. (2023). IL-33's role in the gut immune system: A comprehensive review of its crosstalk and regulation. *Life Sci.* 327: 121868.
- Kelly, C.J., Zheng, L., Campbell, E.L., Saeedi, B., Scholz, C.C., Bayless, A.J., Wilson, K.E., Glover, L.E., Kominsky, D.J., Magnuson, A., Weir, T.L., Ehrentraut, S.F., Pickel, C., Kuhn, K.A., Lanis, J.M., Nguyen, V., Taylor, C.T., and Colgan, S.P. (2015). Crosstalk between Microbiota-Derived Short-Chain Fatty Acids and Intestinal Epithelial HIF Augments Tissue Barrier Function. *Cell Host Microbe* 17(5): 662–671.
- Khan, H.U., Aamir, K., Jusuf, P.R., Sethi, G., Sisinthy, S.P., Ghildyal, R., and Arya, A. (2021). Lauric acid ameliorates lipopolysaccharide (LPS)-induced liver inflammation by mediating TLR4/MyD88 pathway in Sprague Dawley (SD) rats. *Life Sci.* 265: 118750.
- Khan, S., Waliullah, S., Godfrey, V., Khan, M.A.W., Ramachandran, R.A., Cantarel, B.L., Behrendt, C., Peng, L., Hooper, L.V., and Zaki, H. (2020). Dietary simple sugars alter microbial ecology in the gut and promote colitis in mice. *Sci. Transl. Med.* 12(567): eaay6218.
- Kiesler, P., Fuss, I.J., and Strober, W. (2015). Experimental Models of Inflammatory Bowel Diseases. *Cell. Mol. Gastroenterol. Hepatol.* 1(2): 154–170.
- Kikut, J., Drozd, A., Mokrzycka, M., Grzybowska-Chlebowczyk, U., Ziętek, M., and Szczuko, M. (2022). There Is a Differential Pattern in the Fatty Acid Profile in Children with CD Compared to Children with UC. *J. Clin. Med.* 11(9): 2365.
- Kim, J., Ahn, M., Choi, Y., Kang, T., Kim, J., Lee, N.H., Kim, G.O., and Shin, T. (2020). Alpha-Linolenic Acid Alleviates Dextran Sulfate Sodium-

- Induced Ulcerative Colitis in Mice. *Inflammation* 43(5): 1876–1883.
- Kirpich, I.A., Feng, W., Wang, Y., Liu, Y., Barker, D.F., Barve, S.S., and McClain, C.J. (2012). The type of dietary fat modulates intestinal tight junction integrity, gut permeability, and hepatic toll-like receptor expression in a mouse model of alcoholic liver disease. *Alcohol. Clin. Exp. Res.* 36(5): 835–846.
- Kobayashi, T., Siegmund, B., Le Berre, C., Wei, S.C., Ferrante, M., Shen, B., Bernstein, C.N., Danese, S., Peyrin-Biroulet, L., and Hibi, T. (2020). Ulcerative colitis. *Nat. Rev. Dis. Primers* 6(1): 74.
- Koh, A., De Vadder, F., Kovatcheva-Datchary, P., and Backhed, F. (2016). From Dietary Fiber to Host Physiology: Short-Chain Fatty Acids as Key Bacterial Metabolites. *Cell* 165(6): 1332–1345.
- Kong, C., Yan, X., Liu, Y., Huang, L., Zhu, Y., He, J., Gao, R., Kalady, M.F., Goel, A., Qin, H., and Ma, Y. (2021). Ketogenic diet alleviates colitis by reduction of colonic group 3 innate lymphoid cells through altering gut microbiome. *Signal Transduction Targeted Ther.* 6(1): 154.
- Kong, J., Zhang, Z., Musch, M.W., Ning, G., Sun, J., Hart, J., Bissonnette, M., and Li, Y.C. (2008). Novel role of the vitamin D receptor in maintaining the integrity of the intestinal mucosal barrier. *Am. J. Physiol. Gastrointest. Liver Physiol.* 294(1): G208–G216.
- Kono, H., Fujii, H., Ishii, K., Hosomura, N., and Ogiku, M. (2010). Dietary medium-chain triglycerides prevent chemically induced experimental colitis in rats. *Transl. Res.* 155(3): 131–141.
- Koppel, N., Maini Rekdal, V., and Balskus, E.P. (2017). Chemical transformation of xenobiotics by the human gut microbiota. *Science* 356(6344): eaag2770.
- Krause, J.L., Engelmann, B., Schaepe, S.S., Rolle-Kampczyk, U., Jehmlich, N., Chang, H.D., Slanina, U., Hoffman, M., Lehmann, J., Zenclussen, A.C., Herberth, G., von Bergen, M., and Haange, S.B. (2024). DSS treatment does not affect murine colonic microbiota in absence of the host. *Gut Microbes* 16: 2297831.
- Lam, Y.Y., Ha, C.W., Campbell, C.R., Mitchell, A.J., Dinudom, A., Oscarsson, J., Cook, D.I., Hunt, N.H., Caterson, I.D., Holmes, A.J., and Storlien, L.H. (2012). Increased Gut Permeability and Microbiota Change Associate with Mesenteric Fat Inflammation and Metabolic Dysfunction in Diet-Induced Obese Mice. *PLoS One* 7(3): e34233.
- Lam, Y.Y., Ha, C.W., Hoffmann, J.M., Oscarsson, J., Dinudom, A., Mather, T.J., Cook, D.I., Hunt, N.H., Caterson, I.D., Holmes, A.J., and Storlien, L.H. (2015). Effects of dietary fat profile on gut permeability and microbiota and their relationships with metabolic changes in mice. *Obesity* 23(7): 1429–1439.
- Lang, A., Salomon, N., Wu, J.C., Kopylov, U., Lahat, A., Har-Noy, O., Ching, J.Y., Cheong, P.K., Avidan, B., Gamus, D., Kaimakliotis, I., Eliakim, R., Ng, S.C., and Ben-Horin, S. (2015). Curcumin in Combination With Mesalamine Induces Remission in Patients With Mild-to-Moderate Ulcerative Colitis in a Randomized Controlled Trial. *Clin. Gastroenterol. Hepatol.* 13(8): 1444–1449.e1441.
- Laparra, J.M., Alfonso-García, A., Alegría, A., Barberá, R., and Cilla, A. (2015). 7keto-stigmasterol and 7keto-cholesterol induce differential proteome changes to intestinal epithelial (Caco-2) cells. *Food Chem. Toxicol.* 84: 29–36.
- Las Heras, V., Melgar, S., MacSharry, J., and Gahan, C.G.M. (2022). The Influence of the Western Diet on Microbiota and Gastrointestinal Immunity. *Annu. Rev. Food Sci. Technol.* 13: 489–512.
- Le, H.H., Lee, M.-T., Besler, K.R., and Johnson, E.L. (2022). Host hepatic metabolism is modulated by gut microbiota-derived sphingolipids. *Cell Host Microbe* 30(6): 798–808.e797.
- Lee, J.Y., Ye, J., Gao, Z., Youn, H.S., Lee, W.H., Zhao, L., Sizemore, N., and Hwang, D.H. (2003). Reciprocal modulation of Toll-like receptor-4 signaling pathways involving MyD88 and phosphatidylinositol 3-kinase/AKT by saturated and polyunsaturated fatty acids. *J. Biol. Chem.* 278(39): 37041–37051.
- Leon-Coria, A., Kumar, M., Workentine, M., Moreau, F., Surette, M., and Chadee, K. (2021). Muc2 Mucin and Nonmucin Microbiota Confer Distinct Innate Host Defense in Disease Susceptibility and Colonic Injury. *Cell. Mol. Gastroenterol. Hepatol.* 11(1): 77–98.
- Levin, A.D., Wadhwa, V., Leach, S.T., Woodhead, H.J., Lemberg, D.A., Czarina Mendoza-Cruz, A., and Day, A.S. (2011). Vitamin D Deficiency in Children with Inflammatory Bowel Disease. *Dig. Dis. Sci.* 56(3): 830–836.
- Li, F., Han, Y., Cai, X., Gu, M., Sun, J., Qi, C., Goulette, T., Song, M., Li, Z., and Xiao, H. (2020). Dietary resveratrol attenuated colitis and modulated gut microbiota in dextran sulfate sodium-treated mice. *Food Funct.* 11(1): 1063–1073.
- Li, H., Zhu, Y., Zhao, F., Song, S., Li, Y., Xu, X., Zhou, G., and Li, C. (2017). Fish oil, lard and soybean oil differentially shape gut microbiota of middle-aged rats. *Sci. Rep.* 7(1): 826.
- Li, S., Zhuge, A., Wang, K., Lv, L., Bian, X., Yang, L., Xia, J., Jiang, X., Wu, W., Wang, S., Wang, Q., and Li, L. (2021). Ketogenic diet aggravates colitis, impairs intestinal barrier and alters gut microbiota and metabolism in DSS-induced mice. *Food Funct.* 12(20): 10210–10225.
- Li, X., and Li, X. (2020). Obesity Promotes Experimental Colitis by Increasing Oxidative Stress and Mitochondrial Dysfunction in the Colon. *Inflammation* 43(5): 1884–1892.
- Liao, C.M., Zimmer, M.I., and Wang, C.R. (2013). The functions of type I and type II natural killer T cells in inflammatory bowel diseases. *Inflammatory Bowel Dis.* 19(6): 1330–1338.
- Lin, H., An, Y., Hao, F., Wang, Y., and Tang, H. (2016). Correlations of fecal metabolomic and microbiomic changes induced by high-fat diet in the pre-obesity state. *Sci. Rep.* 6(1): 21618.
- Lin, L., and Zhang, J. (2017). Role of intestinal microbiota and metabolites on gut homeostasis and human diseases. *BMC Immunol.* 18(1): 2.
- Lin, Y., Cheng, L., Liu, Y., Wang, Y., Wang, Q., Wang, H.L., Shi, G., Li, J.S., Wang, Q.N., Yang, Q.M., Chen, S., Su, X.L., Yang, Y., Jiang, M., Hu, X., Fan, P., Fang, C., Zhou, Z.G., Dai, L., and Deng, H.X. (2021). Intestinal epithelium-derived BATF3 promotes colitis-associated colon cancer through facilitating CXCL5-mediated neutrophils recruitment. *Mucosal Immunol.* 14(1): 187–198.
- Lissner, D., Schumann, M., Batra, A., Kredel, L.I., Kühl, A.A., Erben, U., May, C., Schulzke, J.D., and Siegmund, B. (2015). Monocyte and M1 Macrophage-induced Barrier Defect Contributes to Chronic Intestinal Inflammation in IBD. *Inflammatory Bowel Dis.* 21(6): 1297–1305.
- Liu, J., Hao, W., He, Z., Kwek, E., Zhao, Y., Zhu, H., Liang, N., Ma, K.Y., Lei, L., He, W.S., and Chen, Z.Y. (2019). Beneficial effects of tea water extracts on the body weight and gut microbiota in C57BL/6J mice fed with a high-fat diet. *Food Funct.* 10(5): 2847–2860.
- Liu, K.Y., Nakatsu, C.H., Jones-Hall, Y., Kozik, A., and Jiang, Q. (2021). Vitamin E alpha- and gamma-tocopherol mitigate colitis, protect intestinal barrier function and modulate the gut microbiota in mice. *Free Radic. Biol. Med.* 163: 180–189.
- Liu, X., Ren, X., Zhou, L., Liu, K., Deng, L., Qing, Q., Li, J., Zhi, F., and Li, M. (2022). Tollip Orchestrates Macrophage Polarization to Alleviate Intestinal Mucosal Inflammation. *J. Crohns. Colitis.* 16(7): 1151–1167.
- Liu, Y., Xiao, H., Wang, Z., Pan, Q., Zhao, X., and Lu, B. (2023). Interactions between dietary cholesterol and intestinal flora and their effects on host health. *Crit. Rev. Food Sci. Nutr.* 65(3): 494–506.
- Liu, Y., Yang, M., Tang, L., Wang, F., Huang, S., Liu, S., Lei, Y., Wang, S., Xie, Z., Wang, W., Zhao, X., Tang, B., and Yang, S. (2022). TLR4 regulates RORγt+ regulatory T-cell responses and susceptibility to colon inflammation through interaction with Akkermansia muciniphila. *Microbiome* 10(1): 98.
- Lloyd-Price, J., Arze, C., Ananthakrishnan, A.N., Schirmer, M., Avila-Pacheco, J., Poon, T.W., Andrews, E., Ajami, N.J., Bonham, K.S., Brislawn, C.J., Casero, D., Courtney, H., Gonzalez, A., Graeber, T.G., Hall, A.B., Lake, K., Landers, C.J., Mallick, H., Plichta, D.R., Prasad, M., Rahnvar, G., Sauk, J., Shungin, D., Vazquez-Baeza, Y., White, R.A. 3rd, Investigators, I., Braun, J., Denson, L.A., Jansson, J.K., Knight, R., Kugathasan, S., McGovern, D.P.B., Petrosino, J.F., Stappenbeck, T.S., Winter, H.S., Clish, C.B., Franzosa, E.A., Vlamakis, H., Xavier, R.J., and Huttenhower, C. (2019). Multi-omics of the gut microbial ecosystem in inflammatory bowel diseases. *Nature* 569(7758): 655–662.
- Łoś-Rycharska, E., Kierasiewicz, Z., and Czerwionka-Szaflarska, M. (2016). Medium chain triglycerides (MCT) formulas in paediatric and allergological practice. *Prz Gastroenterol.* 11(4): 226–231.
- Louis, P., and Flint, H.J. (2017). Formation of propionate and butyrate by the human colonic microbiota. *Environ. Microbiol.* 19(1): 29–41.
- Lu, Q., Yang, M.F., Liang, Y.J., Xu, J., Xu, H.M., Nie, Y.Q., Wang, L.S., Yao, J., and Li, D.F. (2022). Immunology of Inflammatory Bowel Disease: Molecular Mechanisms and Therapeutics. *J. Inflamm. Res.* 15: 1825–1844.
- Lupp, C., Robertson, M.L., Wickham, M.E., Sekirov, I., Champion, O.L., Gaynor, E.C., and Finlay, B.B. (2007). Host-Mediated Inflammation

- Disrupts the Intestinal Microbiota and Promotes the Overgrowth of Enterobacteriaceae. *Cell Host Microbe* 2(2): 119–129.
- Lv, Z., Wang, Y., Yang, T., Zhan, X., Li, Z., Hu, H., Li, T., and Chen, J. (2016). Vitamin A deficiency impacts the structural segregation of gut microbiota in children with persistent diarrhea. *J. Clin. Biochem. Nutr.* 59(2): 113–121.
- Ma, X., Torbenson, M., Hamad, A.R.A., Soloski, M.J., and Li, Z. (2007). High-fat diet modulates non-CD1d-restricted natural killer T cells and regulatory T cells in mouse colon and exacerbates experimental colitis. *Clin. Exp. Immunol.* 151(1): 130–138.
- Machiels, K., Joossens, M., Sabino, J., De Preter, V., Arijs, I., Eeckhaut, V., Ballet, V., Claes, K., Van Immerseel, F., Verbeke, K., Ferrante, M., Verhaegen, J., Rutgeerts, P., and Vermeire, S. (2014). A decrease of the butyrate-producing species *Roseburia hominis* and *Faecalibacterium prausnitzii* defines dysbiosis in patients with ulcerative colitis. *Gut* 63(8): 1275–1283.
- Maldonado-Pereira, L., Schweiss, M., Barnaba, C., and Medina-Meza, I.G. (2018). The role of cholesterol oxidation products in food toxicity. *Food Chem. Toxicol.* 118: 908–939.
- Malmström, V., Shipton, D., Singh, B., Al-Shamkhani, A., Puklavec, M.J., Barclay, A.N., and Powrie, F. (2001). CD134L expression on dendritic cells in the mesenteric lymph nodes drives colitis in T cell-restored SCID mice. *J. Immunol.* 166(11): 6972–6981.
- Maloy, K.J., and Powrie, F. (2011). Intestinal homeostasis and its breakdown in inflammatory bowel disease. *Nature* 474(7351): 298–306.
- Man, S.M., Kaakoush, N.O., and Mitchell, H.M. (2011). The role of bacteria and pattern-recognition receptors in Crohn's disease. *Nat. Rev. Gastroenterol. Hepatol.* 8(3): 152–168.
- Manca, C., Boubertakh, B., Leblanc, N., Deschênes, T., Lacroix, S., Martin, C., Houde, A., Veilleux, A., Flamand, N., Muccioli, G.G., Raymond, F., Cani, P.D., Di Marzo, V., and Silvestri, C. (2020). Germ-free mice exhibit profound gut microbiota-dependent alterations of intestinal endocannabinoid signaling. *J. Lipid Res.* 61(1): 70–85.
- Mandal, S., Godfrey, K.M., McDonald, D., Treuren, W.V., Bjørnholm, J.V., Midtvedt, T., Moen, B., Rudi, K., Knight, R., Brantsæter, A.L., Pedada, S.D., and Eggesbø, M. (2016). Fat and vitamin intakes during pregnancy have stronger relations with a pro-inflammatory maternal microbiota than does carbohydrate intake. *Microbiome* 4(1): 55.
- Martin-Gallausiaux, C., Marinelli, L., Blottiere, H.M., Larraufie, P., and Lapaque, N. (2021). SFA: mechanisms and functional importance in the gut. *Proc. Nutr. Soc.* 80(1): 37–49.
- Martini, E., Krug, S.M., Siegmund, B., Neurath, M.F., and Becker, C. (2017). Mend Your Fences: The Epithelial Barrier and its Relationship With Mucosal Immunity in Inflammatory Bowel Disease. *Cell. Mol. Gastroenterol. Hepatol.* 4(1): 33–46.
- Mascia, C., Maina, M., Chiarpotto, E., Leonarduzzi, G., Poli, G., and Biasi, F. (2010). Proinflammatory effect of cholesterol and its oxidation products on CaCo-2 human enterocyte-like cells: effective protection by epigallocatechin-3-gallate. *Free Radic. Biol. Med.* 49(12): 2049–2057.
- Masnadi Shirazi, K., Nikniaz, Z., Masnadi Shirazi, A., and Rohani, M. (2018). Vitamin A supplementation decreases disease activity index in patients with ulcerative colitis: A randomized controlled clinical trial. *Complement. Ther. Med.* 41: 215–219.
- Mastrodonato, M., Mentino, D., Portincasa, P., Calamita, G., Liquori, G.E., and Ferri, D. (2014). High-fat diet alters the oligosaccharide chains of colon mucins in mice. *Histochem. Cell Biol.* 142(4): 449–459.
- Mayne, C.G., and Williams, C.B. (2013). Induced and natural regulatory T cells in the development of inflammatory bowel disease. *Inflammatory Bowel Dis.* 19(8): 1772–1788.
- Mayr, L., Grabherr, F., Schwarzler, J., Reitmeier, I., Sommer, F., Gehmacher, T., Niederreiter, L., He, G.W., Ruder, B., Kunz, K.T.R., Tymoszuk, P., Hilbe, R., Haschka, D., Feistritz, C., Gerner, R.R., Enrich, B., Przysiecki, N., Seifert, M., Keller, M.A., Oberhuber, G., Sprung, S., Ran, Q., Koch, R., Effenberger, M., Tancevski, I., Zoller, H., Moschen, A.R., Weiss, G., Becker, C., Rosenstiel, P., Kaser, A., Tilg, H., and Adolph, T.E. (2020). Dietary lipids fuel GPX4-restricted enteritis resembling Crohn's disease. *Nat. Commun.* 11(1): 1775.
- Meijer, M.J., Mieremet-Ooms, M.A., van der Zon, A.M., van Duijn, W., van Hogezaand, R.A., Sier, C.F., Hommes, D.W., Lamers, C.B., and Verspaet, H.W. (2007). Increased mucosal matrix metalloproteinase-1, -2, -3 and -9 activity in patients with inflammatory bowel disease and the relation with Crohn's disease phenotype. *Dig. Liver Dis.* 39(8): 733–739.
- Menni, C., Zierer, J., Pallister, T., Jackson, M.A., Long, T., Mohney, R.P., Steves, C.J., Spector, T.D., and Valdes, A.M. (2017). Omega-3 fatty acids correlate with gut microbiome diversity and production of N-carbamylglutamate in middle aged and elderly women. *Sci. Rep.* 7(1): 11079.
- Mielke, L.A., Jones, S.A., Raverdeau, M., Higgs, R., Stefanska, A., Groom, J.R., Misiak, A., Dungan, L.S., Sutton, C.E., Streubel, G., Bracken, A.P., and Mills, K.H. (2013). Retinoic acid expression associates with enhanced IL-22 production by $\gamma\delta$ T cells and innate lymphoid cells and attenuation of intestinal inflammation. *J. Exp. Med.* 210(6): 1117–1124.
- Mirbagheri, S.A., Nezami, B.G., Assa, S., and Hajimahmoodi, M. (2008). Rectal administration of d-alpha tocopherol for active ulcerative colitis: a preliminary report. *World J. Gastroenterol.* 14(39): 5990–5995.
- Mo, Q., Liu, T., Fu, A., Ruan, S., Zhong, H., Tang, J., Zhao, M., Li, Y., Zhu, S., Cai, H., and Feng, F. (2021). Novel Gut Microbiota Patterns Involved in the Attenuation of Dextran Sodium Sulfate-Induced Mouse Colitis Mediated by Glycerol Monolaurate via Inducing Anti-inflammatory Responses. *mBio* 12(5): e0214821.
- Moayyedi, P., Surette, M.G., Kim, P.T., Libertucci, J., Wolfe, M., Onischi, C., Armstrong, D., Marshall, J.K., Kassam, Z., Reinisch, W., and Lee, C.H. (2015). Fecal Microbiota Transplantation Induces Remission in Patients With Active Ulcerative Colitis in a Randomized Controlled Trial. *Gastroenterology* 149(1): 102–109.e106.
- Mottet, C., Uhlig, H.H., and Powrie, F. (2003). Cutting edge: cure of colitis by CD4+CD25+ regulatory T cells. *J. Immunol.* 170(8): 3939–3943.
- Mujico, J.R., Baccan, G.C., Gheorghie, A., Diaz, L.E., and Marcos, A. (2013). Changes in gut microbiota due to supplemented fatty acids in diet-induced obese mice. *Br. J. Nutr.* 110(4): 711–720.
- Murphy, E.A., Velazquez, K.T., and Herbert, K.M. (2015). Influence of high-fat diet on gut microbiota: a driving force for chronic disease risk. *Curr. Opin. Clin. Nutr. Metab.* 18(5): 515–520.
- Muzaki, A.R., Tetlak, P., Sheng, J., Loh, S.C., Setiagani, Y.A., Poidinger, M., Zolezzi, F., Karjalainen, K., and Ruedl, C. (2016). Intestinal CD103+CD11b+ dendritic cells restrain colitis via IFN- γ -induced anti-inflammatory response in epithelial cells. *Mucosal. Immunol.* 9(2): 336–351.
- Nava, P., Koch, S., Laukoetter, M.G., Lee, W.Y., Kolegraff, K., Capaldo, C.T., Beeman, N., Addis, C., Gerner-Smidt, K., Neumaier, I., Skerra, A., Li, L., Parkos, C.A., and Nusrat, A. (2010). Interferon- γ Regulates Intestinal Epithelial Homeostasis through Converging β -Catenin Signaling Pathways. *Immunity* 32(3): 392–402.
- Night, P., Al-Sadi, R., Rawat, M., Guo, S., Watterson, D.M., and Ma, T. (2015). Matrix metalloproteinase 9-induced increase in intestinal epithelial tight junction permeability contributes to the severity of experimental DSS colitis. *Am. J. Physiol.: Gastrointest. Liver Physiol.* 309(12): G988–997.
- Nishida, T., Miwa, H., Shigematsu, A., Yamamoto, M., Iida, M., and Fujishima, M. (1987). Increased arachidonic acid composition of phospholipids in colonic mucosa from patients with active ulcerative colitis. *Gut* 28(8): 1002–1007.
- Ogura, Y., Bonen, D.K., Inohara, N., Nicolae, D.L., Chen, F.F., Ramos, R., Britton, H., Moran, T., Karaliuskas, R., Duerr, R.H., Achkar, J.P., Brant, S.R., Bayless, T.M., Kirschner, B.S., Hanauer, S.B., Nuñez, G., and Cho, J.H. (2001). A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. *Nature* 411(6837): 603–606.
- Okahashi, N., Ueda, M., Matsuda, F., and Arita, M. (2021). Analyses of lipid diversity in gram-negative intestinal bacteria using liquid chromatography–quadrupole time-of-flight mass spectrometry. *Metabolites* 11(4): 197.
- Okamura, T., Hashimoto, Y., Majima, S., Senmaru, T., Ushigome, E., Nakaniishi, N., Asano, M., Yamazaki, M., Takakuwa, H., Hamaguchi, M., and Fukui, M. (2021). Trans Fatty Acid Intake Induces Intestinal Inflammation and Impaired Glucose Tolerance. *Front. Immunol.* 12: 669672.
- Older, E.A., Zhang, J., Ferris, Z.E., Xue, D., Zhong, Z., Mitchell, M.K., Madden, M., Wang, Y., Chen, H., Nagarkatti, P., Nagarkatti, M., Fan, D., Ellermann, M., Li, Y.X., and Li, J. (2024). Biosynthetic enzyme analysis identifies a protective role for TLR4-acting gut microbial sulfonolipids in inflammatory bowel disease. *Nat. Commun.* 15: 9371.

- Oliphant, K., and Allen-Vercoe, E. (2019). Macronutrient metabolism by the human gut microbiome: major fermentation by-products and their impact on host health. *Microbiome* 7(1): 91.
- Olkkonen, V.M., Gylling, H., and Ikonen, E. (2017). Plant sterols, cholesterol precursors and oxysterols: Minute concentrations-Major physiological effects. *J. Steroid Biochem. Mol. Biol.* 169: 4–9.
- Ooi, J.H., Li, Y., Rogers, C.J., and Cantorna, M.T. (2013). Vitamin D Regulates the Gut Microbiome and Protects Mice from Dextran Sodium Sulfate-Induced Colitis. *J. Nutr.* 143(10): 1679–1686.
- Pang, B., Jin, H., Liao, N., Li, J., Jiang, C., and Shi, J. (2021). Vitamin A supplementation ameliorates ulcerative colitis in gut microbiota-dependent manner. *Food Res. Int.* 148: 110568.
- Paone, P., and Cani, P.D. (2020). Mucus barrier, mucins and gut microbiota: the expected slimy partners? *Gut* 69(12): 2232–2243.
- Paramsothy, S., Kamm, M.A., Kaakoush, N.O., Walsh, A.J., van den Bogaerde, J., Samuel, D., Leong, R.W.L., Connor, S., Ng, W., Paramsothy, R., Xuan, W., Lin, E., Mitchell, H.M., and Borody, T.J. (2017). Multidonor intensive faecal microbiota transplantation for active ulcerative colitis: a randomised placebo-controlled trial. *Lancet* 389(10075): 1218–1228.
- Park, B.S., Song, D.H., Kim, H.M., Choi, B.-S., Lee, H., and Lee, J.-O. (2009). The structural basis of lipopolysaccharide recognition by the TLR4-MD-2 complex. *Nature* 458(7242): 1191–1195.
- Pedersen, S.S., Prause, M., Williams, K., Barres, R., and Billestrup, N. (2022). Butyrate inhibits IL-1 β -induced inflammatory gene expression by suppression of NF-kappaB activity in pancreatic beta cells. *J. Biol. Chem.* 298(9): 102312.
- Peterson, C.T., Vaughn, A.R., Sharma, V., Chopra, D., Mills, P.J., Peterson, S.N., and Sivamani, R.K. (2018). Effects of Turmeric and Curcumin Dietary Supplementation on Human Gut Microbiota: A Double-Blind, Randomized, Placebo-Controlled Pilot Study. *J. Evidence-Based Integr. Med.* 23: 2515690X18790725.
- Pigneur, B., and Sokol, H. (2016). Fecal microbiota transplantation in inflammatory bowel disease: the quest for the holy grail. *Mucosal. Immunol.* 9(6): 1360–1365.
- Pillon, N.J., Chan, K.L., Zhang, S., Mejdani, M., Jacobson, M.R., Ducos, A., Bilan, P.J., Niu, W., and Klip, A. (2016). Saturated fatty acids activate caspase-4/5 in human monocytes, triggering IL-1 β and IL-18 release. *Am. J. Physiol. Endocrinol. Metab.* 311(5): E825–e835.
- Pituch-Zdanowska, A., Dembiński, Ł., and Banaszkiewicz, A. (2022). Old but Fancy: Curcumin in Ulcerative Colitis-Current Overview. *Nutrients* 14(24): 5249.
- Prabha, S., Tamoli, S., Raghavamenon, A.C., and Manu, K.A. (2023). Virgin Coconut Oil Alleviates Dextran Sulphate-Induced Inflammatory Bowel Disease and Modulates Inflammation and Immune Response in Mice. *J. Am. Nutr. Assoc.* 43(3): 261–271.
- Progatzky, F., Sangha, N.J., Yoshida, N., McBrien, M., Cheung, J., Shia, A., Scott, J., Marchesi, J.R., Lamb, J.R., Bugeon, L., and Dallman, M.J. (2014). Dietary cholesterol directly induces acute inflammasome-dependent intestinal inflammation. *Nat. Commun.* 5(1): 5864.
- Qiu, P., Ishimoto, T., Fu, L., Zhang, J., Zhang, Z., and Liu, Y. (2022). The Gut Microbiota in Inflammatory Bowel Disease. *Front. Cell. Infect. Microbiol.* 12: 733992.
- Ramanan, D., Tang, M.S., Bowcutt, R., Loke, P., and Cadwell, K. (2014). Bacterial sensor Nod2 prevents inflammation of the small intestine by restricting the expansion of the commensal *Bacteroides vulgatus*. *Immunity* 41(2): 311–324.
- Raphael, I., Nalawade, S., Eagar, T.N., and Forsthuber, T.G. (2015). T cell subsets and their signature cytokines in autoimmune and inflammatory diseases. *Cytokine* 74(1): 5–17.
- Reifen, R., Karlinsky, A., Stark, A.H., Berkovich, Z., and Nyska, A. (2015a). α -Linolenic acid (ALA) is an anti-inflammatory agent in inflammatory bowel disease. *J. Nutr. Biochem.* 26(12): 1632–1640.
- Reifen, R., Levy, E., Berkovich, Z., and Tirosh, O. (2015b). Vitamin A exerts its antiinflammatory activities in colitis through preservation of mitochondrial activity. *Nutrition* 31(11): 1402–1407.
- Resta-Lenert, S., Smitham, J., and Barrett, K.E. (2005). Epithelial dysfunction associated with the development of colitis in conventionally housed *mdr1a*^{-/-} mice. *Am. J. Physiol.: Gastrointest. Liver Physiol.* 289(1): G153–162.
- Richter, H., Gover, O., and Schwartz, B. (2023). Anti-Inflammatory Activity of Black Soldier Fly Oil Associated with Modulation of TLR Signaling: A Metabolomic Approach. *Int. J. Mol. Sci.* 24(13): 10634.
- Rohr, M.W., Narasimhulu, C.A., Rudeski-Rohr, T.A., and Parthasarathy, S. (2020). Negative Effects of a High-Fat Diet on Intestinal Permeability: A Review. *Adv. Nutr.* 11(1): 77–91.
- Rojo, Ó.P., Román, A.L.S., Arbizu, E.A., de la Hera Martínez, A., Sevillano, E.R., and Martínez, A.A. (2006). Serum lipopolysaccharide-binding protein in endotoxemic patients with inflammatory bowel disease. *Inflammatory Bowel Dis.* 13(3): 269–277.
- Röytiö, H., Mokkala, K., Vahlberg, T., and Laitinen, K. (2017). Dietary intake of fat and fibre according to reference values relates to higher gut microbiota richness in overweight pregnant women. *Br. J. Nutr.* 118(5): 343–352.
- Samsami-kor, M., Daryani, N.E., Asl, P.R., and Hekmatdoost, A. (2015). Anti-Inflammatory Effects of Resveratrol in Patients with Ulcerative Colitis: A Randomized, Double-Blind, Placebo-controlled Pilot Study. *Arch. Med. Res.* 46(4): 280–285.
- Samsamikor, M., Daryani, N.E., Asl, P.R., and Hekmatdoost, A. (2016). Resveratrol Supplementation and Oxidative/Anti-Oxidative Status in Patients with Ulcerative Colitis: A Randomized, Double-Blind, Placebo-controlled Pilot Study. *Arch. Med. Res.* 47(4): 304–309.
- Saw, T.Y., Malik, N.A., Lim, K.P., Teo, C.W.L., Wong, E.S.M., Kong, S.C., Fong, C.W., Petkov, J., and Yap, W.N. (2019). Oral supplementation of tocotrienol-rich fraction alleviates severity of ulcerative colitis in mice. *J. Nutr. Sci. Vitaminol.* 65(4): 318–327.
- Schiering, C., Krausgruber, T., Chomka, A., Fröhlich, A., Adelman, K., Wohlfert, E.A., Pott, J., Griseri, T., Bollrath, J., Hegazy, A.N., Harrison, O.J., Owens, B.M.J., Löhning, M., Belkaid, Y., Fallon, P.G., and Powrie, F. (2014). The alarmin IL-33 promotes regulatory T-cell function in the intestine. *Nature* 513(7519): 564–568.
- Seregin, S.S., Golovchenko, N., Schaf, B., Chen, J., Pudlo, N.A., Mitchell, J., Baxter, N.T., Zhao, L., Schloss, P.D., Martens, E.C., Eaton, K.A., and Chen, G.Y. (2017). NLRP6 Protects *Il10*^{-/-} Mice from Colitis by Limiting Colonization of *Akkermansia muciniphila*. *Cell Rep.* 19(4): 733–745.
- Serino, M., Luche, E., Gres, S., Baylac, A., Berge, M., Cenac, C., Waget, A., Klopp, P., Iacovoni, J., Klopp, C., Mariette, J., Bouchez, O., Lluich, J., Ouarne, F., Monsan, P., Valet, P., Roques, C., Amar, J., Bouloumie, A., Theodorou, V., and Burcelin, R. (2011). Metabolic adaptation to a high-fat diet is associated with a change in the gut microbiota. *Gut* 61(4): 543–553.
- Siffledeen, J.S., Siminoski, K., Steinhart, H., Greenberg, G., and Fedorak, R.N. (2003). The Frequency of Vitamin D Deficiency in Adults with Crohn's Disease. *Can. J. Assoc. Gastroenterol.* 17: 391308.
- Simeoli, R., Mattace Raso, G., Pirozzi, C., Lama, A., Santoro, A., Russo, R., Montero-Melendez, T., Berni Canani, R., Calignano, A., Perretti, M., and Meli, R. (2017). An orally administered butyrate-releasing derivative reduces neutrophil recruitment and inflammation in dextran sulphate sodium-induced murine colitis. *Br. J. Pharmacol.* 174(11): 1484–1496.
- Singla, V., Pratap Mouli, V., Garg, S.K., Rai, T., Choudhury, B.N., Verma, P., Deb, R., Tiwari, V., Rohatgi, S., Dhingra, R., Kedia, S., Sharma, P.K., Makharia, G., and Ahuja, V. (2014). Induction with NCB-02 (curcumin) enema for mild-to-moderate distal ulcerative colitis - a randomized, placebo-controlled, pilot study. *J. Crohns. Colitis.* 8: 208–214.
- Souza, C.O., Teixeira, A.A., Biondo, L.A., Silveira, L.S., Calder, P.C., and Rosa Neto, J.C. (2017). Palmitoleic acid reduces the inflammation in LPS-stimulated macrophages by inhibition of NF κ B, independently of PPARs. *Clin. Exp. Pharmacol. Physiol.* 44(5): 566–575.
- Statovci, D., Aguilera, M., MacSharry, J., and Melgar, S. (2017). The Impact of Western Diet and Nutrients on the Microbiota and Immune Response at Mucosal Interfaces. *Front. Immunol.* 8: 838.
- Stephens, M., and von der Weid, P.-Y. (2020). Lipopolysaccharides modulate intestinal epithelial permeability and inflammation in a species-specific manner. *Gut microbes* 11(3): 421–432.
- Sugihara, K., Morhardt, T.L., and Kamada, N. (2018). The Role of Dietary Nutrients in Inflammatory Bowel Disease. *Front. Immunol.* 9: 3183.
- Sugimoto, K., Hanai, H., Tozawa, K., Aoshi, T., Uchijima, M., Nagata, T., and Koide, Y. (2002). Curcumin prevents and ameliorates trinitrobenzene sulfonic acid-induced colitis in mice. *Gastroenterology* 123(6): 1912–1922.
- Sun, J., Qiao, Y., Qi, C., Jiang, W., Xiao, H., Shi, Y., and Le, G.-w. (2016). High-

- fat-diet-induced obesity is associated with decreased anti-inflammatory Lactobacillus reuteri sensitive to oxidative stress in mouse Peyer's patches. *Nutrition* 32(2): 265–272.
- Sun, Q., Zhang, S., Liu, X., Huo, Y., Su, B., and Li, X. (2020). Effects of a probiotic intervention on Escherichia coli and high-fat diet-induced intestinal microbiota imbalance. *Appl. Microbiol. Biotechnol.* 104: 1243–1257.
- Suzuki, T., and Hara, H. (2010). Dietary fat and bile juice, but not obesity, are responsible for the increase in small intestinal permeability induced through the suppression of tight junction protein expression in LETO and OLETF rats. *Nutr. Metab.* 7: 1–17.
- Tahan, G., Aytac, E., Aytakin, H., Gunduz, F., Dogusoy, G., Aydin, S., Tahan, V., and Uzun, H. (2011). Vitamin E has a dual effect of anti-inflammatory and antioxidant activities in acetic acid-induced ulcerative colitis in rats. *Can. J. Surg.* 54(5): 333–338.
- Tajika, M., Matsuura, A., Nakamura, T., Suzuki, T., Sawaki, A., Kato, T., Hara, K., Ookubo, K., Yamao, K., Kato, M., and Muto, Y. (2004). Risk factors for vitamin D deficiency in patients with Crohn's disease. *J. Gastroenterol.* 39(6): 527–533.
- Tedelind, S., Westberg, F., Kjerrulf, M., and Vidal, A. (2007). Anti-inflammatory properties of the short-chain fatty acids acetate and propionate: a study with relevance to inflammatory bowel disease. *World J. Gastroenterol.* 13(20): 2826–2832.
- Tian, Y., Nichols, R.G., Cai, J., Patterson, A.D., and Cantorna, M.T. (2018). Vitamin A deficiency in mice alters host and gut microbial metabolism leading to altered energy homeostasis. *J. Nutr. Biochem.* 54: 28–34.
- Tomas, J., Mulet, C., Saffarian, A., Cavin, J.B., Ducroc, R., Regnault, B., Kun Tan, C., Duszka, K., Burcelin, R., Wahli, W., Sansonetti, P.J., and Pédrón, T. (2016). High-fat diet modifies the PPAR- γ pathway leading to disruption of microbial and physiological ecosystem in murine small intestine. *Proc. Natl. Acad. Sci. U.S.A.* 113(40): E5934–e5943.
- Tong, L.C., Wang, Y., Wang, Z.B., Liu, W.Y., Sun, S., Li, L., Su, D.F., and Zhang, L.C. (2016). Propionate Ameliorates Dextran Sodium Sulfate-Induced Colitis by Improving Intestinal Barrier Function and Reducing Inflammation and Oxidative Stress. *Front. Pharmacol.* 7: 253.
- Tulkens, J., Vergauwen, G., Van Deun, J., Geeurickx, E., Dhondt, B., Lippens, L., De Scheerder, M.A., Miinalainen, I., Rappu, P., De Geest, B.G., Vandecasteele, K., Laukens, D., Vandekerckhove, L., Denys, H., Vandesompele, J., De Wever, O., and Hendrix, A. (2020). Increased levels of systemic LPS-positive bacterial extracellular vesicles in patients with intestinal barrier dysfunction. *Gut* 69(1): 191–193.
- Usami, M., Komurasaki, T., Hanada, A., Kinoshita, K., and Ohata, A. (2003). Effect of γ -linolenic acid or docosahexaenoic acid on tight junction permeability in intestinal monolayer cells and their mechanism by protein kinase C activation and/or eicosanoid formation. *Nutrition* 19(2): 150–156.
- van der Hee, B., and Wells, J.M. (2021). Microbial Regulation of Host Physiology by Short-chain Fatty Acids. *Trends Microbiol.* 29(8): 700–712.
- Van der Sluis, M., De Koning, B.A., De Bruijn, A.C., Velcich, A., Meijerink, J.P., Van Goudoever, J.B., Büller, H.A., Dekker, J., Van Seuningen, I., Renes, I.B., and Einerhand, A.W. (2006). Muc2-deficient mice spontaneously develop colitis, indicating that MUC2 is critical for colonic protection. *Gastroenterology* 131(1): 117–129.
- van der Wulp, M.Y.M., Verkade, H.J., and Groen, A.K. (2013). Regulation of cholesterol homeostasis. *Mol. Cell. Endocrinol.* 368(1): 1–16.
- Van Klinken, B.J., Van der Wal, J.W., Einerhand, A.W., Büller, H.A., and Dekker, J. (1999). Sulphation and secretion of the predominant secretory human colonic mucin MUC2 in ulcerative colitis. *Gut* 44(3): 387–393.
- Vatanen, T., Kostic, A.D., d'Hennezel, E., Siljander, H., Franzosa, E.A., Yassour, M., Kolde, R., Vlamakis, H., Arthur, T.D., Hamalainen, A.M., Peet, A., Tillmann, V., Uibo, R., Mokurov, S., Dorshakova, N., Ilonen, J., Virtanen, S.M., Szabo, S.J., Porter, J.A., Lahdesmaki, H., Huttenhower, C., Gevers, D., Cullen, T.W., Knip, M., DIABIMMUNE Study Group, and Xavier, R.J. (2016). Variation in Microbiome LPS Immunogenicity Contributes to Autoimmunity in Humans. *Cell* 165(4): 842–853.
- Venema, K. (2010). Role of gut microbiota in the control of energy and carbohydrate metabolism. *Curr. Opin. Clin. Nutr. Metab. Care* 13(4): 432–438.
- Vignali, D.A., Collison, L.W., and Workman, C.J. (2008). How regulatory T cells work. *Nat. Rev. Immunol.* 8(7): 523–532.
- Villamor, E., and Fawzi, W.W. (2000). Vitamin A Supplementation: Implications for Morbidity and Mortality in Children. *J. Infect. Dis.* 182(Suppl 1): S122–S133.
- Walker, A., Pfitzner, B., Harir, M., Schaubek, M., Calasan, J., Heinzmann, S.S., Turav, D., Rattai, T., Endesfelder, D., Castell, W.Z., Haller, D., Schmid, M., Hartmann, A., and Schmitt-Kopplin, P. (2017). Sulfonolipids as novel metabolite markers of Alistipes and Odoribacter affected by high-fat diets. *Sci. Rep.* 7(1): 11047.
- Wan, Y., Yuan, J., Li, J., Li, H., Zhang, J., Tang, J., Ni, Y., Huang, T., Wang, F., Zhao, F., and Li, D. (2020). Unconjugated and secondary bile acid profiles in response to higher-fat, lower-carbohydrate diet and associated with related gut microbiota: A 6-month randomized controlled-feeding trial. *Clin. Nutr.* 39(2): 395–404.
- Wang, D.Q. (2007). Regulation of intestinal cholesterol absorption. *Annu. Rev. Physiol.* 69: 221–248.
- Wang, J., Huang, N., Xiong, J., Wei, H., Jiang, S., and Peng, J. (2018). Caprylic acid and nonanoic acid upregulate endogenous host defense peptides to enhance intestinal epithelial immunological barrier function via histone deacetylase inhibition. *Int. Immunopharmacol.* 65: 303–311.
- Wang, Y., An, Y., Ma, W., Yu, H., Lu, Y., Zhang, X., Wang, Y., Liu, W., Wang, T., and Xiao, R. (2020). 27-Hydroxycholesterol contributes to cognitive deficits in APP/PS1 transgenic mice through microbiota dysbiosis and intestinal barrier dysfunction. *J. Neuroinflammation* 17(1): 199.
- Watson, H., Mitra, S., Croden, F.C., Taylor, M., Wood, H.M., Perry, S.L., Spencer, J.A., Quirke, P., Toogood, G.J., Lawton, C.L., Dye, L., Loadman, P.M., and Hull, M.A. (2018). A randomised trial of the effect of omega-3 polyunsaturated fatty acid supplements on the human intestinal microbiota. *Gut* 67(11): 1974–1983.
- Wei, M., Gao, X., Liu, L., Li, Z., Wan, Z., Dong, Y., Chen, X., Niu, Y., Zhang, J., and Yang, G. (2020). Visceral Adipose Tissue Derived Exosomes Exacerbate Colitis Severity via Pro-inflammatory MiRNAs in High Fat Diet Fed Mice. *ACS Nano* 14(4): 5099–5110.
- Wilson, M.S., Ramalingam, T.R., Rivollier, A., Shenderov, K., Mentink-Kane, M.M., Madala, S.K., Cheever, A.W., Artis, D., Kelsall, B.L., and Wynn, T.A. (2011). Colitis and intestinal inflammation in IL10-/- mice results from IL-13R α 2-mediated attenuation of IL-13 activity. *Gastroenterology* 140(1): 254–264.
- Wolf, B.J., Titurri, R.V., Almeida, C.F., Le Nours, J., Bhowruth, V., Johnson, D., Uldrich, A.P., Hsu, F.F., Brigl, M., Besra, G.S., Rossjohn, J., Godfrey, D.I., and Brenner, M.B. (2015). Identification of a Potent Microbial Lipid Antigen for Diverse NKT Cells. *J. Immunol.* 195(6): 2540–2551.
- Wolf, P.G., Gaskins, H.R., Ridlon, J.M., Freels, S., Hamm, A., Goldberg, S., Petrilli, P., Schering, T., Vergis, S., Gomez-Perez, S., Yazici, C., Braunschweig, C., Mutlu, E., and Tussing-Humphreys, L. (2020). Effects of taurocholic acid metabolism by gut bacteria: a controlled feeding trial in adult African American subjects at elevated risk for colorectal cancer. *Contemp. Clin. Trials Commun.* 19: 100611.
- Wolters, M., Ahrens, J., Romani-Pérez, M., Watkins, C., Sanz, Y., Benítez-Páez, A., Stanton, C., and Günther, K. (2019). Dietary fat, the gut microbiota, and metabolic health—A systematic review conducted within the MyNewGut project. *Clin. Nutr.* 38(6): 2504–2520.
- Wu, D., Ye, X., Linhardt, R.J., Liu, X., Zhu, K., Yu, C., Ding, T., Liu, D., He, Q., and Chen, S. (2021). Dietary pectic substances enhance gut health by its polycapillary component: A review. *Compr. Rev. Food Sci. Food Saf.* 20(2): 2015–2039.
- Wu, Q., Luo, Y., Lu, H., Xie, T., Hu, Z., Chu, Z., and Luo, F. (2024). The Potential Role of Vitamin E and the Mechanism in the Prevention and Treatment of Inflammatory Bowel Disease. *Foods* 13(6): 898.
- Wunderlich, C.M., Ackermann, P.J., Ostermann, A.L., Adams-Quack, P., Vogt, M.C., Tran, M.L., Nikolajev, A., Waisman, A., Garbers, C., Theurich, S., Mauer, J., Hövelmeyer, N., and Wunderlich, F.T. (2018). Obesity exacerbates colitis-associated cancer via IL-6-regulated macrophage polarisation and CCL-20/CCR-6-mediated lymphocyte recruitment. *Nat. Commun.* 9(1): 1646.
- Wynn, T.A., Chawla, A., and Pollard, J.W. (2013). Macrophage biology in development, homeostasis and disease. *Nature* 496(7446): 445–455.
- Xie, R., Sun, Y., Wu, J., Huang, S., Jin, G., Guo, Z., Zhang, Y., Liu, T., Liu, X., Cao, X., Wang, B., and Cao, H. (2018). Maternal High Fat Diet Alters Gut Microbiota of Offspring and Exacerbates DSS-Induced Colitis in Adulthood. *Front. Immunol.* 9: 2608.
- Xu, G., Guan, L., Sun, J., and Chen, Z.-Y. (2009). Oxidation of cholesterol and β -sitosterol and prevention by natural antioxidants. *J. Agric. Food*

- Chem. 57(19): 9284–9292.
- Xu, G., Sun, J., Liang, Y., Yang, C., and Chen, Z.-Y. (2011). Interaction of fatty acids with oxidation of cholesterol and β -sitosterol. *Food Chem.* 124(1): 162–170.
- Xu, X., Ocansey, D.K.W., Pei, B., Zhang, Y., Wang, N., Wang, Z., and Mao, F. (2023). Resveratrol alleviates DSS-induced IBD in mice by regulating the intestinal microbiota-macrophage-arginine metabolism axis. *Eur. J. Med. Res.* 28(1): 319.
- Yan, C., Huang, S.H., Ding, H.F., Kwek, E., Liu, J.H., Chen, Z.X., Ma, K.Y., and Chen, Z.Y. (2023). Adverse effect of oxidized cholesterol exposure on colitis is mediated by modulation of gut microbiota. *J. Hazard. Mater.* 459: 132057.
- Yan, C., Kwek, E., Ding, H.-F., He, Z., Ma, K.Y., Zhu, H., and Chen, Z.-Y. (2022). Dietary oxidized cholesterol aggravates chemically induced murine colon inflammation and alters gut microbial ecology. *J. Agric. Food Chem.* 70(41): 13289–13301.
- Yang, L., Sakandar, H.A., Sun, Z., and Zhang, H. (2021). Recent advances of intestinal microbiota transmission from mother to infant. *J. Funct. Foods* 87: 104719.
- Yang, W., Zhao, P., Li, X., Guo, L., and Gao, W. (2022). The potential roles of natural plant polysaccharides in inflammatory bowel disease: A review. *Carbohydr. Polym.* 277: 118821.
- Yao, J., Lu, Y., Zhi, M., Hu, P., Wu, W., and Gao, X. (2017). Dietary n-3 polyunsaturated fatty acids ameliorate Crohn's disease in rats by modulating the expression of PPAR- γ /NFAT. *Mol. Med. Rep.* 16(6): 8315–8322.
- Yao, Y., Cai, X., Fei, W., Ye, Y., Zhao, M., and Zheng, C. (2022). The role of short-chain fatty acids in immunity, inflammation and metabolism. *Crit. Rev. Food Sci. Nutr.* 62(1): 1–12.
- Ye, Z., Xu, Y.-J., and Liu, Y. (2021). Influences of dietary oils and fats, and the accompanied minor content of components on the gut microbiota and gut inflammation: A review. *Trends Food Sci. Technol.* 113: 255–276.
- Yip, J.L.K., Balasuriya, G.K., Spencer, S.J., and Hill-Yardin, E.L. (2021). The Role of Intestinal Macrophages in Gastrointestinal Homeostasis: Heterogeneity and Implications in Disease. *Cell. Mol. Gastroenterol. Hepatol.* 12(5): 1701–1718.
- Zaph, C., Du, Y., Saenz, S.A., Nair, M.G., Perrigoue, J.G., Taylor, B.C., Troy, A.E., Kobuley, D.E., Kastelein, R.A., Cua, D.J., Yu, Y., and Artis, D. (2008). Commensal-dependent expression of IL-25 regulates the IL-23–IL-17 axis in the intestine. *J. Exp. Med.* 205(10): 2191–2198.
- Zeissig, S., Bürgel, N., Günzel, D., Richter, J., Mankertz, J., Wahnschaffe, U., Kroesen, A.J., Zeitz, M., Fromm, M., and Schulzke, J.D. (2007). Changes in expression and distribution of claudin 2, 5 and 8 lead to discontinuous tight junctions and barrier dysfunction in active Crohn's disease. *Gut* 56(1): 61–72.
- Zeng, W.-P. (2013). 'All things considered': transcriptional regulation of T helper type 2 cell differentiation from precursor to effector activation. *Immunology* 140(1): 31–38.
- Zhang, X., Coker, O.O., Chu, E.S., Fu, K., Lau, H.C.H., Wang, Y.X., Chan, A.W.H., Wei, H., Yang, X., Sung, J.J.Y., and Yu, J. (2021). Dietary cholesterol drives fatty liver-associated liver cancer by modulating gut microbiota and metabolites. *Gut* 70(4): 761–774.
- Zhang, Y.-M., and Rock, C.O. (2008). Membrane lipid homeostasis in bacteria. *Nat. Rev. Microbiol.* 6(3): 222–233.
- Zhang, Z., Xue, Z., Yang, H., Zhao, F., Liu, C., Chen, J., Lu, S., Zou, Z., Zhou, Y., and Zhang, X. (2021). Differential effects of EPA and DHA on DSS-induced colitis in mice and possible mechanisms involved. *Food Funct.* 12(4): 1803–1817.
- Zhao, D., Cai, C., Chen, Q., Jin, S., Yang, B., and Li, N. (2020). High-fat diet promotes DSS-induced ulcerative colitis by downregulated FXR expression through the TGFB pathway. *Biomed. Res. Int.* 2020: 3516128.
- Zheng, D., Liwinski, T., and Elinav, E. (2020). Interaction between microbiota and immunity in health and disease. *Cell Res.* 30(6): 492–506.
- Zheng, L., Kelly, C.J., Battista, K.D., Schaefer, R., Lanis, J.M., Alexeev, E.E., Wang, R.X., Onyiah, J.C., Kominsky, D.J., and Colgan, S.P. (2017). Microbial-Derived Butyrate Promotes Epithelial Barrier Function through IL-10 Receptor-Dependent Repression of Claudin-2. *J. Immunol.* 199(8): 2976–2984.
- Zheng, X.F., Hong, Y.X., Feng, G.J., Zhang, G.F., Rogers, H., Lewis, M.A., Williams, D.W., Xia, Z.F., Song, B., and Wei, X.Q. (2013). Lipopolysaccharide-induced M2 to M1 macrophage transformation for IL-12p70 production is blocked by *Candida albicans* mediated up-regulation of EB13 expression. *PLoS One* 8(5): e63967.
- Zhong, V.W., Van Horn, L., Cornelis, M.C., Wilkins, J.T., Ning, H., Carnethon, M.R., Greenland, P., Mentz, R.J., Tucker, K.L., Zhao, L., Norwood, A.F., Lloyd-Jones, D.M., and Allen, N.B. (2019). Associations of Dietary Cholesterol or Egg Consumption With Incident Cardiovascular Disease and Mortality. *JAMA* 321(11): 1081–1095.
- Zhou, S., Wang, Y., Jacoby, J.J., Jiang, Y., Zhang, Y., and Yu, L.L. (2017). Effects of Medium- and Long-Chain Triacylglycerols on Lipid Metabolism and Gut Microbiota Composition in C57BL/6J Mice. *J. Agric. Food Chem.* 65(31): 6599–6607.
- Zhu, F., Zheng, J., Xu, F., Xi, Y., Chen, J., and Xu, X. (2021). Resveratrol alleviates dextran sulfate sodium-induced acute ulcerative colitis in mice by mediating PI3K/Akt/VEGFA pathway. *Frontiers in pharmacology* 12: 693982.